

# Sublethal effects of pesticide residues differ between strains of captive Grey partridge: consequences in terms of rearing conditions.

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1	Sublethal effects of pesticide residues differ between strains of captive Grey partridge:
2	consequences in terms of rearing conditions
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23	ABSTRACT
24	Over the last 50 years, farmland bird populations have declined steeply in Europe and North
24	America. Reintroduction or reinforcement for populations unable to self-maintain are popular
23	America. Reinforduction of reinforcement for populations unable to sen-maintain are populat

management tools to overcome extinction risk through captive rearing and release. However, 26 27 released birds tend to have lower survival rates than their wild conspecifics. Here, we aimed to mimic the diet shift that Grey partridge (*Perdix perdix*) encounters once released (poultry 28 29 food to natural grain) and compared (i) differences in responses to this transition depending on strain (wild or farm) and (ii) the consequences of ingesting organic or conventional grains 30 on individual body mass, haematocrit and behavioural traits (anti-predator responses, activity, 31 32 exploration) that are likely critical for post-release survival. Our experimental procedure involved 40 farm-strain and 40 wild-strain partridges fed with commercial poultry food 33 during early life then with wheat and corn for 3 months, from either organic or conventional 34 (clopyralid residues) agriculture. The results suggest that the haematocrit and body mass 35 index of farm-strain partridges were lower at the end of the experiment, probably due to diet 36 shift. Birds fed conventional grains (with clopyralid residues) exhibited lower flight duration 37 and flight initiation distance, and were more likely to translocate by running rather than 38 flying. The findings suggest that, when rearing partridges of farm-strain origin, birds fed 39 organic grains experience better post-release survival than birds fed conventional grains 40 containing pesticide residues. 41

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43 **KEYWORDS:** captivity, conservation, strain, *Perdix perdix*, pesticide residues

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### 45 **1. INTRODUCTION**

Over the last 50 years, farmland bird populations have declined by 20% in Europe (Burns et al. 2021) and 74% in North America (Stanton et al. 2018). The main causes of decline are related to agricultural intensification, including habitat loss, seasonal shifts in cultivation practices, and increased use of agrochemicals (Robinson and Sutherland 2002; Newton 2004). Reintroductions after strong decline (e.g. the process of rearing animals in captivity and

releasing them into the wild) or reinforcement for populations unable to self-maintain are two 51 management tools that have become increasingly popular in the past 30 years to overcome 52 extinction risk, even on a local scale (Seddon et al. 2007). Reinforcing wild populations has 53 also been commonly used in game birds for hunting purposes, leading to game farm 54 implementation, which bred originally wild-strain birds in captivity for decades (Buner et al. 55 2011). However, released farm-strain individuals such as common pheasant (Musil and 56 57 Connelly 2009), chukar partridge (Yolcu et al. 2016) and grey partridge (Parish and Sotherton 2007) tend to have lower survival rates than their wild conspecifics (Sokos et al. 2009). 58 Understanding why farm-strain birds are less fit to persist in wild environments may help us 59 to improve reintroduction projects through changes in rearing conditions. 60

Captive environments are known to impair genetic diversity, lead to genetic drift, and 61 enhance adaptation to captivity, all of which can be detrimental when such individuals are 62 63 released into the wild (Frankham 2008; Keller et al. 2012). Indeed, many studies have reported behavioural deficiencies in farm-strain birds that impair survival (Rymešová et al. 64 65 2012, 2013; Homberger et al. 2014; Yolcu et al. 2016), especially in regard to excessive boldness (Madden and Whiteside 2014), inhibited escape responses (Pérez et al. 2010), lower 66 group vigilance (Rantanen et al. 2010b) and poor anti-predator defences (Håkansson and 67 68 Jensen 2008; Rantanen et al. 2010a). Upon release, animals have to cope with risk-taking, foraging and novel environment discovery (Réale et al. 2007). Fleeing quickly in life-69 threatening situations, and showing a high level of exploration and activity to find food, 70 sexual partners and territory, should collectively increase fitness (Sih et al. 2004; Smith and 71 72 Blumstein 2008; Homberger et al. 2013). Moreover, farm-strain birds display higher body mass, lower glycogen reserves and lower oxidative capacity than wild ones as a result of 73 74 unlimited access to high-nutrient food in captivity (Putaala and Hissa 1995, 1998). In addition to making them less fit, captivity accustoms their metabolism to conditions that sharply 75

diverge from those that await them in nature (i.e. food unpredictability and quality, variable 76 diet, vulnerability to predators and/or diseases). Food quality during captivity may thus also 77 play a role in post-release survival and the success of the reintroduction process (Madden et 78 al. 2020). Traditionally, farm-strain birds are fed with highly-digestible commercial poultry 79 food (Liukkonen-Anttila et al. 1999, 2000) including genetically modified soya, wheat and 80 corn that differs from their natural diet (in terms of energy supply, digestibility and quality). 81 However, whether in processed or unprocessed form, grains used in farm-strain bird diets may 82 contain pesticide residues used in conventional agriculture (Kumar et al. 2019). Even small 83 quantities of pesticides in food may cause sublethal effects when eaten over an extended time 84 (Parween et al. 2016; Moreau et al. 2021). In game birds, exposure to pesticide treatments are 85 known to deteriorate the immune system (Lopez-Antia et al. 2015b) and haematocrit (Lopez-86 Antia et al. 2015a), but also induce dysfunctions of metabolic pathways associated with the 87 88 ability to store fat ( Lopez-Antia et al. 2013; Moreau et al. 2021), and interfere with the endocrine system (Lopez-Antia et al. 2015c; Khalil et al. 2017). The latter results in 89 behavioural changes in general activity (Moreau et al. 2021), risk-taking behaviour (Kobiela 90 91 et al. 2015) and vigilance (de Faria et al. 2018), which may have positive or negative effects on survival (i.e. foraging, reproduction, predation) depending on environmental context 92 (Smith and Blumstein 2008). 93

Therefore, strain (wild or farm) and food quality may affect physiological or behavioural traits in released individuals, which could alter the fitness and long-term demographics of population though simultaneous effects that have received limited attention (but see Homberger et al. 2013, 2015 and 2021). In the present study, we used Grey partridge (*Perdix perdix*), an iconic game bird of European farmlands that is currently declining, as a test case (Sotherton et al. 2010; Aebischer and Ewald 2012; Bro 2016). Several million birds are annually reared and released in Europe, including ~2 million in France (Bro 2016;

Madden 2021). Mortality rates of released farm-strain partridges are very high at 60–80% 101 102 (Parish and Sotherton 2007; Buner and Schaub 2008; Rymešová et al. 2013; Bro 2016; Homberger et al. 2021). Although many studies have investigated morphological, behavioural 103 104 and physiological deficiencies, as well as post-release survival of farm-strain grey partridges (Putaala and Hissa 1995, 1998; Rantanen et al. 2010a, 2010b; Keller et al. 2012; Homberger 105 et al. 2013, 2015, 2021; Rymešová et al. 2013), none have explored the potential interactions 106 107 between strain (wild or farm) and sublethal effects of pesticides. We thus aimed to mimic the 108 diet shift that partridges encounter once released (poultry food vs. natural grain) to investigate (i) differences in responses to this transition depending on strain, and (ii) the consequences of 109 ingesting organic or conventional grains on physiology and behaviour traits that are likely 110 critical for post-release survival. Our experimental procedure involved 8-month-old grey 111 partridges, either bred in captivity for 60 generations (farm strain) or 2 generations (wild 112 113 strain). Partridges were fed from birth with poultry food. At the beginning of the experiment, they were fed with natural grain grown by either organic or conventional agriculture. 114 115 Individual conditions were monitored throughout captivity and several behavioural traits were 116 also quantified for each bird (see Methods for details). We predicted that birds that had been in captivity the longest (farm strain) would lose weight and display a lower haematocrit in 117 comparison to wild-strain birds due to diet shift. Furthermore, we predicted that birds fed 118 conventional grains would be even more affected in terms of physiology and behaviour due to 119 potential pesticide residue ingestion. 120

121

- 122 2. MATERIALS AND METHODS
- 123 **2.1 Experimental design**

124 The study took place in a commercial game farm in Southwest France, and lasted 3 months,125 from December 2019 to March 2020. Tests were performed on 80 8-month-old captive-born

grey partridges; 40 were from a game farm strain (60 to 80 generations in captivity, 'farm-126 strain' hereafter) and 40 were from F3 generations of wild trapped birds (two generations in 127 captivity, 'wild-strain' hereafter). From birth to the beginning of the experiment 128 (corresponding to the diet shift from poultry food to natural grain), all birds had the same 129 experience, living in two large pens depending on their strain (100 m  $\times$  10 m  $\times$  4 m) at the 130 game farm. They were fed ground feed mixture for game birds (STARGIB entretien; see ESM 131 132 1 for composition) mainly made from imported conventional soya bean, maize, wheat and sunflower, likely containing pesticide residues (Klüche M. et al. 2020). Before 133 134 experimentation commenced, each bird was sexed (according to the presence of secondary sexual characters) and fitted with an alphanumeric metal ring. 135

In December 2019, four experimental groups of 20 partridges were established 136 randomly, including 10 males and 10 females, each placed in a grassy holding pen (100 m  $\times$ 137  $10 \text{ m} \times 4 \text{ m}$ ) including a game bird feeder, several drinkers equally spaced in the pen, and two 138 sheets of metal providing shelter. The first group comprised 20 wild-strain partridges fed ad 139 libitum with grains obtained from harvests of certified organic crops (i.e. purchased from 140 141 organic producers), composed of half wheat and half corn. The second group of 20 wild-strain partridges was fed ad libitum with grains purchased from producers in conventional 142 agriculture (i.e. for which different pesticides were used during the cropping season) also 143 composed of half wheat and half corn. Similarly, the third and the fourth groups were 144 composed of 20 farm-strain partridges each, either fed ad libitum with organic or 145 conventional wheat and corn. 146

A previous study showed that conventional grains contain pesticide residuals whereas organic grains do not, and that the energy content for both types is equivalent (Moreau et al. 2021). In the present study, only conventional grown wheat had detectable and quantifiable pesticide residues (analysed by GIRPA, a referenced laboratory based at Beaucouze, Loire

Atlantique, France). We detected 0.014 mg/kg clopyralid, above the limit of quantification set 151 at 0.010 mg/kg. Clopyralid is an organochlorine herbicide that is considered relatively non-152 toxic to terrestrial and aquatic animals, although available data concerning toxicity to birds 153 154 are scarce (EFSA 2018). No pesticide residues were detected in organic grains. Birds were under constant surveillance, with water and food provided each day by the farmer from the 155 18th of December (week 0) to the 10th of March (week 12). All groups were kept under a 156 natural light cycle, and 11 partridges died during experimentation (4 in the wild-strain organic 157 group, 3 in the wild-strain conventional group, 1 in the farm-strain organic group, and 2 in the 158 farm-strain conventional group; Fisher's exact test; p = 0.57). 159

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# 2.2 Monitored parameters

During the experiment, birds in each group were monitored to assess (i) their physiological condition through haematocrit, which reflects nutritional state and physiological stress (Ots et al. 1998), and body mass index, (which reflects energy capital accumulated by feeding (Peig and Green 2009), and (ii) their behaviour in relation to anti-predator responses, activity, and exploration (Réale et al. 2007; de Faria et al. 2018).

166 2.2.1 Physiological conditions; haematocrit and body mass index

Morphological measurements and blood samples were taken the day before release, while in 167 168 their pens, in December (week 0) and after 3 months of food type (week 12). To assess haematocrit, a blood sample was taken from the brachial vein using a sterile needle (Ø 0.06 169 mm) and heparinised micro-capillary tubes (10 µL). Capillary tubes were then centrifuged at 170 171 5000 rpm for 5 min immediately afterwards to determine haematocrit levels; the proportion of the tube filled with red blood cells was divided by the total sample volume in the capillary 172 tube. (Biard et al. 2015). Length was measured with an electronic calliper to the nearest 0.1 173 174 mm. In order to compare the body mass index of grey partridges, right and left tarsus lengths were measured twice with a calliper (accuracy 0.1 mm) at the beginning of the experiment 175

(week 0), since in adulthood tarsus lengths remain stable. Body mass was recorded at week 0
and week 12 to the nearest 1 g using a spring scale (Pesola 500 g). From body mass and mean
tarsus length (right and left), we evaluated the body mass index with the scaled mass index
(Peig and Green 2009; Moreau et al. 2021).

180 2.2.2 Behavioural tests

A set of behavioural tests were performed in week 9 during three consecutive days with 181 identical weather (from the 18<sup>th</sup> to the 20<sup>th</sup> of February); 21 birds were tested on day 1, 27 on 182 day 2, and 27 on day 3. Each evening, partridges of different groups were randomly caught, 183 placed in cardboard boxes (five birds per box, 66 cm  $\times$  38 cm  $\times$  17.5 cm), and left inside for 184 the night under controlled conditions (7-11°C). Catching birds the night before the 185 experiment enabled us to decrease the capture stress that can affect behavioural tests. The 186 187 following day, each bird was processed using the same sequence, which consisted in (i) the cage test and (ii) the open-field test. The cage test consisted of measuring flight initiation 188 189 distance (FID) and escape attempt strategy (as birds could not escape) in a cage outside. The open-field test consisted of measuring activity, exploration, escape strategy and duration of 190 191 escape in an open-field test. The behaviour of each bird was always recorded by the same 192 observers. After the full series of measurements, birds were released into their pens.

193 *2.2.3 Cage test* 

Partridges were placed by a single operator in a square, wire cage ( $46 \text{ cm} \times 32 \text{ cm} \times 36 \text{ cm}$ ) in a ploughed field. Another operator was hidden behind a hedge, 100 m from the cage, and they watched the bird's behaviour through binoculars. After 5 min of acclimatisation, the hidden operator approached the bird. The FID, the distance at which the individual begins to fly when a predator, or human, approaches (Frid & Dill, 2002), was estimated as the distance at which the bird reacted (tried to run or fly inside the cage) using the straight-line approach of the operator. The escape attempt strategy (running or flying) of the bird was also recorded. Approaches towards birds were performed by the same operator to standardise approach speed and operator dress, thus limiting detectability bias. After the test, birds were caught by the first operator, placed in a cardboard box ( $66 \text{ cm} \times 38 \text{ cm} \times 17.5 \text{ cm}$ ) and brought directly to the open-field test in a nearby building ( $5 \text{ m} \times 5.5 \text{ m} \times 1.50 \text{ m}$ , 7–11°C, 30.5 lux).

205 2.2.4 Open-field test

A grid was set on the floor of the building that included 110 numbered squares, each  $50 \times 50$ 206 cm. Partridges were brought in a cardboard box (66 cm  $\times$  38 cm  $\times$  17.5 cm) and given 3 min 207 208 for acclimatisation to the box. The operator, hidden in a corner behind a sheet, then opened the cardboard box to let the bird out. The exploration behaviour of the partridge was assessed 209 by counting the number of different squares visited during 7 min. Activity, considered any 210 behaviour except resting, as measured as the number of times each square was visited. At the 211 end of exploration and activity assessment, a stuffed fox ( $40 \text{ cm} \times 80 \text{ cm}$ ) on castors, hidden 212 with the operator, was launched towards birds at a speed of  $\sim 1$  m/s. The responses to this 213 stimulus were then assessed as (i) their escape strategy (running or flying) and (ii) their 214 duration of flight (recorded with a chronometer). After the test, partridges were caught with a 215 landing net and returned to their cage (see Fig. 1). 216

217

# 2.3 Statistical analysis

We analysed the effects of strains and treatments on the physiological conditions of birds at 218 219 three different times. First, at the beginning (week 0) and at the end (week 12) of the experiment, we tested whether strain (wild vs. farm), food type (organic vs. conventional), 220 and their combination affected haematocrit and body mass index, accounting for sex 221 differences using analysis of variance (ANOVA). Although no effect of food type was 222 involved at week 0 since all birds were fed with the very same food until that date, food effect 223 224 from the start of the experiment was included in the model to test for possible initial sampling bias. A third model investigated whether trends existed for haematocrits or body mass index 225

between week 0 and week 12, including the effects of strain, sex, and food nested within 226 227 strain. Pairwise post-hoc comparison tests between the four groups were tested using Bonferroni correction in the case where interactions between strain and food type were 228 229 significant. We chose to nest the effects of food and strain because at the beginning of the experiment we already had a strain effect (see the results below). Similarly, we analysed the 230 effects of strain and food type on behavioural traits, including their interactions and sex 231 differences. Activity and exploration were square root transformed before analysis. Both 232 escape strategy during cage and open-field tests could be in the form of flight or running, 233 hence we used a binomial modelling framework in the logistic regression model and escape 234 235 strategy (0 for running, 1 for flying) as a response variable. Finally, for birds that flew away during the open-field test, duration of flight was also square root transformed before analysis. 236 The normality of residuals and the distribution of residuals against fitted values were checked 237 238 for each model. Effect of blood sample hours on haematocrit variation was also checked at week 0 using simple linear model (LM). For all tests, p-values were calculated at the 5% 239 240 significance level. All statistical tests were performed using R software (v. 3.5.0, R Core 241 Team 2018) and the smatr package (Warton et al. 2012) for scale mass index computation (Peig and Green 2009). Results are presented as means  $\pm$  standard error (SE). 242

243 **2.4 Ethical approval** 

All experiments complied with French laws on animal experimentation, and all experimental
protocols were approved by the Committee of Animal Experimentations of the Deux-Sèvres
French Department (APA FIS#9465-201703101551625).

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#### **3. RESULTS**

Numbers of monitored partridges differed depending on the test undertaken (this is specifiedon each figure). Concerning physiological condition comparison, 65 partridges were

monitored (11 deaths at the end of the experiment and four without all data). Concerning
behavioural tests, 73 partridges were monitored because at the time of the test, seven
partridges were already dead.

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## 3.1 Partridge physiological conditions

Time of blood samples did not affect haematocrit variation (LM  $\chi^2 = 1.77$ , df = 1, P = 0.19). 255 256 Farm-strain partridges had slightly but significantly lower haematocrits than wild birds at the 257 onset of the experiment, with haematocrits of birds in the conventional food type group significantly lower than in the other groups (i.e. highlighting a sampling bias, Fig. 2.a, Table 1). 258 259 At week 12, farm-strain partridges still had lower haematocrits (Fig. 2.a). Haematocrits of all four groups increased with both food types (significant intercepts when comparing week 0 and 12) 260 with a significant effect of sex (Fig. 2.a, Table 1). Indeed, haematocrits of males (mean  $\pm$  SE at 261 week 0; 42.96  $\pm$  3.17% at week 12; 46.91  $\pm$  3.53%) increased more than those of females (43.33  $\pm$ 262 3.37% and 44.85  $\pm$  3.27%; Fig. 2.a, Table 1). Thus, there was no statistically significant effect of 263 264 food type on haematocrit.

Conversely, the body mass index of farm-strain partridges was significantly higher than that of wild birds at the onset of the experiment (Fig. 2.b, Table 1), and female body mass index was higher that of males (week 0,  $388.8 \pm 31.8$  g for females,  $357.5 \pm 28.7$  g for males). At week 12 a significant sex effect was evident, with female body mass index remaining higher than that of males (week 12,  $365.1 \pm 37.1$  g for females,  $331.2 \pm 22.8$  g for males). In addition, body mass index decreased during the food type experiment only for farm-strain partridges regardless of food type, whereas this index stayed stable for wild birds (Fig. 2.b, Table 1).

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#### 3.2 Partridge behaviour

Strain and food type affected behavioural traits in different ways (Table 2). During the cage test, farm-strain partridges fed conventional grains showed lower FID ( $46.1 \pm 22.2$  m) than other birds (farm-strain birds fed organic grains =  $63.1 \pm 13.2$  m; wild-strain birds fed organic

grains =  $56.7 \pm 16.9$  m; wild-strain birds fed conventional grains =  $65.2 \pm 18.9$  m; Fig. 3.a). 276 Moreover, strain and food type interacted significantly to impact escape strategy; farm-strain 277 partridges fed conventional grains were less likely to escape by flying in comparison to other 278 279 groups (Table 2, Table 3). Concerning the open-field test, farm-strain partridges were less active (Table 2, Fig. 3.b) and less likely to try to escape by flying in response to the stuffed fox in 280 comparison with wild-strain birds (Table 2, Table 3). Finally, for birds that escaped by flying, 281 282 duration of flight was higher for wild-strain partridges fed organic grains, and farm-strain birds that consumed conventional grains displayed the shortest flight duration among the four groups 283 (Table 2, Fig. 3.c). 284

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#### 286 4. DISCUSSION

In this study, we compared the effects of captive-reared bird strain (farm vs. wild) and 287 interactions with potential sublethal effects of low doses of pesticide ingestion through food 288 (organic vs. conventional grains), though only clopyralid was detected. Regardless of food 289 type, the body mass index of farm-strain birds was higher at the onset of the experiment, and 290 291 thus decreased as the experiment progressed, while that of wild-strain birds remained stable. Conversely, haematocrit of farm-strain partridges was lower at the onset of the experiment but 292 293 increased over time for all groups. We also found that farm-strain partridges had lower activity and tended to flee by running rather than flying during the open-field test. Finally, we 294 found an interaction between strain and food type in behavioural traits involved in anti-295 296 predator responses, with farm-strain birds fed conventional grains (containing clopyralid 297 residues) showing lower FID and shorter flight duration, and they were less likely to try to escape by flying during cage tests than other groups. Such an interaction between strain and 298 food type resulted in the worst performance for farm-strain birds fed conventional grains. 299 These results suggest that these birds could be disadvantaged in the wild. 300

Captive-bred grey partridges are traditionally fed *ad libitum* using feeders containing 301 commercial poultry food; however, when released, these birds have to cope with an 302 unpredictable and strikingly different type of food, based on winter grains (Perkins et al. 303 304 2007). Our experimental design somewhat mimicked this transition as diet of birds shifted from poultry food to natural wheat and corn grains at the onset of the experiment. As 305 predicted, farm-strain partridges showed higher body mass than wild-strain birds at the 306 beginning of the experiment, highlighting morphological differences between the two strains. 307 308 Indeed, captive-reared birds are often larger than their wild counterparts as a result of unlimited access to high-nutrient food over several generations of captivity (Putaala and Hissa 309 1995, 1998). Regardless of treatment type, we found that the body mass index of farm-strain 310 birds decreased, which could suggest that those birds may be less inclined to properly 311 assimilate new/unfamiliar food. Indeed, captivity adaptation could have led to shortened 312 313 intestines, caeca and gizzards of our farm-strain partridges, in comparison with wild birds (Putaala and Hissa 1995; Liukkonen- Anttila et al. 2002), although we did not examine the 314 315 digestive systems of our birds in order to keep them alive. These organs play a role in food 316 digestion and assimilation, and the larger they are, the more effective they are at processing fibrous food such as natural grains. As expected, wild-strain partridges seemed to cope better 317 318 with this dietary change, based on their relatively stable body mass index, consistent with 319 previous studies (Putaala and Hissa 1995). An alternative hypothesis may explain the decrease in body mass of farm-strain partridges (i.e. the shift to a smaller group and/or a different 320 group composition). Indeed, the stress responses and behaviour of individuals depends on the 321 group to which they belong, as reported previously for body condition (Creel et al. 2013; 322 Homberger et al. 2015). Testing this hypothesis would have required performing enough 323 replicates for each group, which was not feasible here due to space constraints in the game 324 farm. 325

Haematocrit is often positively correlated with body condition as it reflects nutritional 326 state and physiological stress, although extrapolating from an individual's state should be 327 done with caution (Ots et al. 1998; Fair et al. 2007). However, in the current study, we found 328 329 the opposite trend between fat storage and red blood cells. Indeed, at the beginning of the experiment, farm-strain partridges (i.e. those with the higher body condition) had lower 330 haematocrit than wild birds. Although we detected significant sampling bias at the onset of 331 the experiment, haematocrit increased over the course of the study for all four groups. This 332 overall increase in haematocrit may be linked to the mating period, which begins in February 333 for partridges, since haematocrit can change according to hormones (Fair et al. 2007). This 334 hypothesis was corroborated by the haematocrits of males, which were higher those of 335 females at the end of the experiment, highlighting differences between sex, even though 336 haematocrit increased for all birds (Fair et al. 2007). 337

338 We also found that farm-strain partridges were less active and preferred to flee by running rather than by flying during open-field tests. Physiological and morphological 339 340 adaptations due to long-term captivity (e.g., higher body mass for farm-strain partridges at week 0) may have induced behavioural changes in our farm-strain partridges. For example, 341 farm-strain partridges may have weaker endurance due to smaller breast muscles, lower 342 343 glycogen content, and consequently lower release and/or production of glucocorticoids in pectoral muscles (Putaala and Hissa 1995). Moreover, they may to have lower excretion of 344 glucocorticoids as a response to stress, which can affect anti-predator responses (Homberger 345 et al., 2013, 2014, 2015). As animals selected over many generations for high reproductive 346 rates, with food provided ad libitum, one may expect a decrease in energy-demanding 347 behaviours for farm-strain birds, such as escape by flying (Beilharz et al. 1993; Schutz and 348 Jensen 2001). In addition, being isolated from predators may result in lacking appropriate 349 anti-predator responses (Pérez et al. 2010; Rantanen et al. 2010b). Upon release, animals have 350

to cope with risk-taking, foraging and novel environment discovery (Réale et al. 2007). 351 Fleeing quickly in life-threatening situations (e.g., by flying rather than running), and 352 displaying a high level of activity to find food, sexual partners and territories are expected to 353 354 increase fitness (Sih et al. 2004; Smith and Blumstein 2008; Homberger et al. 2013). Based on our results, our farm-strain partridges could therefore be expected to suffer higher mortality 355 rates than their wild counterparts after release (Rantanen et al. 2010a; Rymešová et al. 2012, 356 357 2013; Homberger et al. 2014). Interestingly, Homberger et al. (2021) found that bolder birds (i.e., those with higher activity and lower tonic immobility responses) survived longer after 358 release despite negative associations postulated and repeatedly reported between proactivity, 359 360 bold behaviour, survival and longevity (Dammhahn et al., 2018; Smith & Blumstein 2007). Under fluctuating environmental conditions or changing needs at different stages of the life 361 cycle, the advantages and disadvantages of a particular behaviour type may change or even be 362 363 reversed (Sih et al., 2004). Individuals can acquire their own behavioural strategies and adjust behaviour according to environmental context, making it difficult to predict their probability 364 of survival based solely on captive experiments. 365

We also detected an interaction between strain and food type, since farm-strain birds 366 fed conventional grains showed lower capacity to detect (i.e., lower FID) and escape a 367 predator in the field (i.e., running instead of flying). Many studies have described sublethal 368 adverse effects of pesticides on behaviour, caused by neurotoxicity (Mitra et al. 2011; Moreau 369 et al. 2022, submitted). For example, organomercury fungicides act mainly through the central 370 nervous system, resulting in disruption to risk-taking behaviour (Kobiela et al. 2015); 371 abamectin insecticides may inhibit GABA receptors and cause environment perception 372 disorder (de Faria et al. 2018); neonicotinoids impact acetylcholine esterase activity, leading 373 to a decrease in foraging activity and ultimately to starvation and body weight loss (Eng et al. 374 2017, 2019). Although clopyralid is considered relatively non-toxic to birds (EFSA 2018), 375

some long-term studies have reported weight loss and damage to stomach, liver and kidney at
15 mg/kg per day for mammals, specifically rats and rabbits (Hayes et al. 1984; EFSA 2018).
Our current results suggest that chronic ingestion of clopyralid residues may induce
neurotoxicity and alter anti-predator responses of farm-strain partridges, whereas wild-strain
partridges seemed to be unaffected by conventional grain ingestion.

381

#### 382 5. CONCLUSION

Currently, most commercial game farms breed partridges from farm-strain birds and feed 383 them commercial poultry food (Sokos et al. 2009; Homberger et al. 2014). Our results suggest 384 385 that game farms produce birds that have lower fitness (i.e., less likely to survive and/or reproduce) when released into the wild compared with wild strains, and this is aggravated by 386 food type, (i.e. when fed conventional grains containing pesticide residues). Our results 387 therefore suggest that feeding captive-bred individuals from game farms with organic grains 388 may, to some extent, compensate for this disadvantage, and may therefore contribute to 389 improving post-release survival rates. This strategy may be preferred by partridge farmers 390 who may be more inclined to change the food they provide their partridges rather than 391 completely changing their breeding strategy. 392

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394

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401

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#### 410 8. REFERENCES

Aebischer NJ, Ewald JA, Game & Wildlife Conservation Trust, Fordingbridge, UK (2012)
The grey partridge in the UK: population status, research, policy and prospects. Anim
Biodivers Conserv 35:353–362. https://doi.org/10.32800/abc.2012.35.0353

Beilharz RG, Luxford BG, Wilkinson JL (1993) Quantitative genetics and evolution: Is our
understanding of genetics sufficient to explain evolution? J Anim Breed Genet 110:161–170.
https://doi.org/10.1111/j.1439-0388.1993.tb00728.x

Biard C, Monceau K, Motreuil S, Moreau J (2015) Interpreting immunological indices: The
importance of taking parasite community into account. An example in blackbirds Turdus
merula. Methods Ecol Evol 6:960–972. https://doi.org/10.1111/2041-210X.12371

420 Bro E (2016) La Perdrix grise. Biologie, écologie, gestion et conservation, Biotope. Mèze,
421 France

Buner F, Browne S, Aebischer N (2011) Experimental assessment of release methods for the
re-establishment of a red-listed galliform, the Grey Partridge (Perdix perdix). Biol Conserv
144:593–601. https://doi.org/10.1016/j.biocon.2010.10.017

Buner F, Schaub M (2008) How do different releasing techniques affect the survival of
reintroduced grey partridges *Perdix perdix*?. Wildlife Biol 14:26–35.
https://doi.org/10.2981/0909-6396(2008)14[26:HDDRTA]2.0.CO;2

Burns F, Eaton MA, Burfield IJ, Klvaňová A, Šilarová E, Staneva A, Gregory RD (2021)
Abundance decline in the avifauna of the European Union reveals cross - continental
similarities in biodiversity change. Ecol Evol 11:16647-16660.
https://doi.org/10.1002/ece3.8282

Creel S, Dantzer B, Goymann W, Rubenstein DR, Boonstra R (2013) The ecology of stress:
Effects of the social environment. Funct. Ecol., 27:66–80. <u>https://doi.org/10.1111/j.1365-</u>
<u>2435.2012.02029</u>.

Dammhahn M., Dingemanse NJ, Niemelä PT, Réale D (2018) Pace-of-life syndromes: A
framework for the adaptive integration of behaviour, physiology and life history. Behav. Ecol.
and Sociobiol. 72: 20132645. https://doi.org/10.1007/s00265-018-2473-y

de Faria DBG, Montalvão MF, Chagas TQ, Araújo APC, Souza JM, Mendes BO, Rodrigues
ASL, Malafaia G (2018) Behavioral changes in Japanese quails exposed to predicted
environmentally relevant abamectin concentrations. Sci Total Environ 636:1553–1564.
https://doi.org/10.1016/j.scitotenv.2018.04.293

Eng ML, Stutchbury BJM, Morrissey CA (2017) Imidacloprid and chlorpyrifos insecticides
impair migratory ability in a seed-eating songbird. Sci Rep 7. https://doi.org/10.1038/s41598017-15446-x

- Eng ML, Stutchbury BJM, Morrissey CA (2019) A neonicotinoid insecticide reduces fueling
  and delays migration in songbirds. Science 365:1177–1180.
  https://doi.org/10.1126/science.aaw9419
- 448 E.F.S.A (2018) Conclusion on the peer review of the pesticide risk assessment of the active
- substance clopyralid. EFSA Journal 16:5389, 21 pp. https://doi.org/10.2903/j.efsa.2018.5389
- 450 Fair J, Whitaker S, Pearson B (2007) Sources of variation in haematocrit in birds. Ibis
  451 149:535–552. https://doi.org/10.1111/j.1474-919X.2007.00680.x
- 452 Frankham R (2008) Genetic adaptation to captivity in species conservation programs. Mol
- 453 Ecol 17:325–333. https://doi.org/10.1111/j.1365-294X.2007.03399.x
- Håkansson J, Jensen P (2008) A longitudinal study of antipredator behaviour in four
  successive generations of two populations of captive red junglefowl. Appl Anim Behav Sci
  114:409–418. https://doi.org/10.1016/j.applanim.2008.04.003
- Hayes WC, Smith FA, John JA, Rao KS (1984) Teratologic Evaluation of 3,6Dichloropicolinic Acid in Rats and Rabbits. Toxicol Sci 4:91–97.
  https://doi.org/10.1093/toxsci/4.1.91
- Homberger B, Jenni L, Duplain J, Lanz M, Schaud M (2014) Food unpredictability in early
  life increases survival of captive grey partridges (*Perdix perdix*) after release into the wild.
  Biol Conserv 177:134–141. https://doi.org/10.1016/j.biocon.2014.06.023
- Homberger B, Jenni-Eiermann S, Jenni L (2015) Distinct responses of baseline and stressinduced corticosterone levels to genetic and environmental factors. Gen. Comp. Endocrin.
  210:46-54 https://doi.org/10.1016/j.ygcen.2014.09.020.
- Homberger B, Jenni-Eiermann S, Roulin A, Jenni L (2013) The impact of pre- and post-natal
  contexts on immunity, glucocorticoids and oxidative stress resistance in wild and

- domesticated grey partridges. Funct Ecol 27:1042–1054. https://doi.org/10.1111/13652435.12092
- 470 Homberger B, Korner-Nievergelt F, Jenni-Eiermann S, Duplain J, Lanz M, Jenni L (2021)
- 471 Integrating behaviour, physiology and survival to explore the outcome of reintroductions: a
  472 case study of grey partridge. Anim Behav 176:145–156.
  473 https://doi.org/10.1016/j.anbehav.2021.04.005
- Keller LF, Biebach I, Ewing SR, Hoeck PEA (2012) The genetics of reintroductions:
  inbreeding and genetic drift. In: Reintroduction Biology, John Wily & Sons Ltd (Eds), pp 360–
  394. https://doi.org/10.1002/9781444355833.ch11
- Khalil SR, Awad A, Mohammed HH (2017) Behavioral response and gene expression
  changes in fipronil-administered male Japanese quail (*Coturnix japonica*). Environ Pollut
  223:51–61. https://doi.org/10.1016/j.envpol.2016.12.027
- Klüche M, Bikker P, van Egmond H.J, Hoffmans Y, Mol J.G.J (2020) Potential presence of
  pesticide
- residues in feed, requiring single residue methods (SRM). Wageningen Food Safety Research,
  WFSR report, 46 pp.
- Kobiela ME, Cristol DA, Swaddle JP (2015) Risk-taking behaviours in zebra finches affected
  by mercury exposure. Anim Behav 103:153–160.
  https://doi.org/10.1016/j.anbehav.2015.02.024
- Kumar V, Kumar P, Singh J (2019) An introduction to contaminants in agriculture and
  environment. In: Contaminants in agriculture and environment: health risks and remediation,
  Agro Environ Media Agriculture and Environmental Science Academy (Eds), Haridwar,
  India, pp 1–8

- Liukkonen-Anttila T, Putaala A, Hissa R (1999) Does shifting from a commercial to a natural
  diet affect the nutritional status of hand-reared grey partridges Perdix perdix? Wildl Biol
  5:147–156. https://doi.org/10.2981/wlb.1999.019
- Liukkonen-Anttila T, Saartoala R, Hissa R (2000) Impact of hand-rearing on morphology and
  physiology of the capercaillie (Tetrao urogallus). Comp Biochem Physiol A Mol Integr
  Physiol 125:211–221. https://doi.org/10.1016/S1095-6433(99)00174-9
- Liukkonen Anttila T, Uimaniemi L, Orell M, Lumme J (2002) Mitochondrial DNA 497 variation and the phylogeography of the grey partridge (Perdix perdix) in Europe: from 498 Pleistocene history populations. Biol 499 to present day J Evol 15:971–982. 500 https://doi.org/10.1046/j.1420-9101.2002.00460.x
- Lopez-Antia A, Ortiz-Santaliestra ME, Camarero PR, et al (2015c) Assessing the Risk of
  Fipronil-Treated Seed Ingestion and Associated Adverse Effects in the Red-Legged Partridge.
  Environ Sci Technol 49:13649–13657. https://doi.org/10.1021/acs.est.5b03822
- 504 Lopez-Antia A, Ortiz-Santaliestra ME, García-de Blas E, et al (2015b) Adverse effects of 505 thiram-treated seed ingestion on the reproductive performance and the offspring immune 506 function of the red-legged partridge. Environ Toxicol Chem 34:1320–1329. 507 https://doi.org/10.1002/etc.2925
- Lopez-Antia A, Ortiz-Santaliestra ME, Mougeot F, Mateo R (2013) Experimental exposure of
  red-legged partridges (*Alectoris rufa*) to seeds coated with imidacloprid, thiram and
  difenoconazole. Ecotoxicology 22, 125–138. https://doi.org/10.1007/s10646-012-1009-x
- 511 Lopez-Antia A, Ortiz-Santaliestra ME, Mougeot F, Mateo R (2015a) Imidacloprid-treated 512 seed ingestion has lethal effect on adult partridges and reduces both breeding investment and
- offspring immunity. Environ Res 136:97–107. https://doi.org/10.1016/j.envres.2014.10.023

- 514 Madden J, Santilli F, Whiteside M (2020) The welfare of game birds destined for release into
- the wild: a balance between early life care and preparation for future natural hazards. Anim
- 516 Welf 29:1–18. https://doi.org/10.7120/09627286.29.1.001
- 517 Madden JR (2021) How many gamebirds are released in the UK each year? Eur J Wildl Res
  518 67:72. https://doi.org/10.1007/s10344-021-01508-z
- Madden JR, Whiteside MA (2014) Selection on behavioural traits during 'unselective'
  harvesting means that shy pheasants better survive a hunting season. Anim Behav 87:129–
- 521 135. https://doi.org/10.1016/j.anbehav.2013.10.021
- 522 Mitra A, Chatterjee C, Mandal FB (2011) Synthetic chemical pesticides and their effects on
  523 birds. Res J Environ Toxicol 5:81–96. https://doi.org/10.3923/rjet.2011.81.96
- 524 Moreau J, Monceau K, Crépin M, Tochon FD, Mondet C, Fraikin M, Teixeira M, Bretagnolle
- 525 V (2021) Feeding partridges with organic or conventional grain triggers cascading effects in
- 526 life-history traits. Environ Pollut 278:116851. https://doi.org/10.1016/j.envpol.2021.116851
- 527 Musil DD, Connelly JW (2009) Survival and reproduction of pen reared vs translocated
- wild pheasants Phasianus colchicus. Wildl Biol 15:80–88. https://doi.org/10.2981/07-049
- Newton I (2004) The recent declines of farmland bird populations in Britain: An appraisal of
  causal factors and conservation actions. Ibis 146:579–600. https://doi.org/10.1111/j.1474919X.2004.00375.x
- Ots I, MurumÄgi A, Hõrak P (1998) Haematological health state indices of reproducing Great
  Tits: methodology and sources of natural variation. Funct Ecol 12:700–707.
  https://doi.org/10.1046/j.1365-2435.1998.00219.x

Parish DMB, Sotherton NW (2007) The fate of released captive-reared grey partridges Perdix
perdix: implications for reintroduction programmes. Wildl Biol 13:140–149.
https://doi.org/10.2981/0909-6396(2007)13[140:TFORCG]2.0.CO;2

Parween T, Jan S, Mahmooduzzafar S, Fatma T, Siddiqui ZH (2016) Selective effect of
pesticides on plant—A review. Crit Rev Food Sci Nutr 56:160–179.
https://doi.org/10.1080/10408398.2013.787969

- Peig J, Green AJ (2009) New perspectives for estimating body condition from mass/length
  data: the scaled mass index as an alternative method. Oikos 118:1883–1891.
  https://doi.org/10.1111/j.1600-0706.2009.17643.x
- Pérez JA, Alonso ME, Prieto R, Bartolomé D, Gaudioso DVR (2010) Influence of the
  breeding system on the escape response of red-legged partridges (Alectoris rufa). Poult Sci
  89:5–12. https://doi.org/10.3382/ps.2009-00170
- Perkins AJ, Anderson G, Wilson JD (2007) Seed food preferences of granivorous farmland
  passerines. Bird Study 54:46–53. https://doi.org/10.1080/00063650709461455
- Putaala A, Hissa R (1995) Effects of hand-rearing on physiology and anatomy in the grey
  partridge. Wildl Biol 1:27–31. https://doi.org/10.2981/wlb.1995.006
- Putaala A, Hissa R (1998) Breeding dispersal and demography of wild and hand-reared grey
  partridges Perdix perdix in Finland. Wildl Biol 4:137–145.
  https://doi.org/10.2981/wlb.1998.016
- Rantanen EM, Buner F, Riordan P, Sotherton N, Macdonald DW (2010a) Habitat preferences 554 and survival in wildlife reintroductions: an ecological trap in reintroduced grey partridges: 555 Ecological traps in wildlife reintroductions. J Appl Ecol 47:1357-1364. 556 https://doi.org/10.1111/j.1365-2664.2010.01867.x 557

- Rantanen EM, Buner F, Riordan P, Sotherton N, Macdonald DW (2010b) Vigilance, time
  budgets and predation risk in reintroduced captive-bred grey partridges Perdix perdix. Appl
  Anim Behav Sci 127:43–50. https://doi.org/10.1016/j.applanim.2010.08.002
- Réale D, Reader SM, Sol D, McDougall PT, Dingemanse NJ (2007) Integrating animal
  temperament within ecology and evolution. Biol Rev 82:291–318.
  https://doi.org/10.1111/j.1469-185X.2007.00010.x
- Robinson RA, Sutherland WJ (2002) Post-war changes in arable farming and biodiversity in
  Great Britain. J Appl Ecol 39:157–176
- Rymešová D, Šmilauer P, Šálek M (2012) Sex- and age-biased mortality in wild Grey
  Partridge Perdix perdix populations. Ibis 154:815–824. https://doi.org/10.1111/j.1474919X.2012.01259.x
- Rymešová D, Tomášek O, Šálek M (2013) Differences in mortality rates, dispersal distances
  and breeding success of commercially reared and wild grey partridges in the Czech
  agricultural landscape. Eur J Wildl Res 59:147–158. https://doi.org/10.1007/s10344-0120659-6
- Schutz K, Jensen P (2001) Effects of resource allocation on behavioural strategies: a
  comparison of red junglefowl (Gallus gallus) and two domesticated breeds of poultry.
  Ethology 107:753–765. https://doi.org/10.1046/j.1439-0310.2001.00703.x
- 576 Seddon PJ, Armstrong DP, Maloney RF (2007) Developing the science of reintroduction
- 577 biology. Conserv Biol 21:303–312. https://doi.org/10.1111/j.1523-1739.2006.00627.x
- Sih A, Bell A, Johnson JC (2004) Behavioral syndromes: an ecological and evolutionary
  overview. Trends Ecol Evol 19:372–378. https://doi.org/10.1016/j.tree.2004.04.009

- 580 Smith BR, Blumstein DT (2008) Fitness consequences of personality: a meta-analysis. Behav
- 581 Ecol 19:448–455. https://doi.org/10.1093/beheco/arm144
- Sokos C, Birtsas P, Tsachalidis E (2009) The aims of galliforms release and choice of
  techniques. Wildl Biol 14:412–422. https://doi.org/10.2981/0909-6396-14.4.412
- 584 Sotherton NW, Aebischer NJ, Ewald JA (2010) The conservation of the Grey Partridge. In:
- Maclean N (ed) Silent summer: the state of wildlife in Britain and Ireland. Cambridge
  University Press, Cambridge, pp 319–336
- 587 Stanton RL, Morrissey CA, Clark RG (2018) Analysis of trends and agricultural drivers of
- farmland bird declines in North America: A review. Agric Ecosyst Environ 254:244–254.
- 589 https://doi.org/10.1016/j.agee.2017.11.028
- 590 Sutherland W, Armstrong D, Butchart S, Earnhardt J, Ewen J, Jamieson I, Jones C, Lee R,
- 591 Newbery P, Nichols J, Parker K, Sarrazin F, Seddon P, Shah N, Tatayah V (2010) Standards
- for documenting and monitoring bird reintroduction projects. Conserv. Lett. 3:229–235.
- 593 https://doi.org/10.1111/j.1755-263X.2010.00113.x
- Warton DI, Duursma RA, Falster DS and Taskinen S (2012) Smatr 3 an R package for
  estimation and inference about allometric lines. Methods Ecol. Evol. 3:257-259.
  https://doi.org/10.1111/j.2041-210X.2011.00153.x
- 597 Yolcu Hİ, Aslan A, Serttaş A, Saribaşak H, Uysal H, Çobanoğlu A (2016) Monitoring the
- survival rate of released chukars: a case study inElmalı Cedar Forest, Antalya, Turkey. Turk J
- 599 Zool 40:272–278. https://doi.org/10.3906/zoo-1506-5

Table 1 Effects of strain (wild vs. farm), sex and food type (organic vs. conventional) on the physiological condition of grey partridges (haematocrit and body mass index) at the beginning and end of the food experiment (week 0 before food type and week 12), and differences at 12 weeks based on the combined effect of food and strain. Wild birds fed organic grains served as a reference in models.  $\beta \pm$ SE values were extracted from minimal models including only significant factors. Farm, farm-strain; Conv., conventional. Significant effects are in bold.

		Beginning	g of tl	ne experir	nent (week 0)	tt (week 0) End of the experiment (week 12)					Difference between week 12 and week 0			
Response	Fixed	F	df	р	$\beta\pm SE$	F	df	р	$\beta\pm SE$	F	df	р	$\beta\pm SE$	
variables	factors													
Haematocrit					Intercept =				Intercept =				Intercept = 2.30 ± 0.84,	
					$44.42\pm0.66$				$46.08\pm0.91$				<i>p</i> <0.01	
	Strain	6.99	1	<0.01	Farm = -2.06	6.34	1	<0.01	Farm = -1.63	0.01	1	0.98		
					± 0.78				± 1.15					
	Sex	0.25	1	0.62		6.21	1	<0.01	Males = 2.05	10.23	1	<0.01	Males = $2.52 \pm 0.76$	
									± 0.82					
	Food	1.54	1	0.22		0.91	1	0.35						

	Strain	8.10	1	<0.01	Farm Conv.	0.27	2	0.61		2.45	2	0.09	
	x Food				= -4.19 ± 1.47								
Body mas	5				Intercept =				Intercept =				Intercept = $-6.03 \pm 6.46$
index					$375.70\pm5.81$				$376.26\pm8.73$				
	Strain	14.44	1	<0.001	Farm = 25.52	0.24	1	0.63		26.30	1	<0.001	Farm = -31.61 ± 8.15
					± 6.80								
	Sex	20.83	1	<0.001	Male = -31.19	19.44	1	<0.001	Male = -34.05	0.53	1	0.53	
					± 6.83				± 7.88				
	Food	1.37	1	0.25		0.42	1	0.52					
	Strain	2.36	1	0.13		2.50	2	0.12		0.19	2	0.83	
	x Food												

Table 2 Effects of strain (wild vs. farm), sex and food type (organic vs. conventional) on partridge behaviour during cage and open-field tests. Linear models were conducted for FID and square root transformed activity, exploration, and duration of flight. Logistic regression was conducted for escape strategy. Wild birds fed organic grains served as a reference in models.  $\beta \pm$  SE values were extracted from minimal models including only significant factors. Significant effects are in bold.

Tests	Response	Fixed factors	F	df	р	$\beta\pm SE$
						Intercept = $57.73 \pm 4.71$
		Strain	1.09	1	0.30	
	FID	Food	1.98	1	0.16	
	(n = 73)	Sex	0.24	1	0.62	
_		Strain x Food	8.97	2	<0.01	Farm-strain conventional =
Cage test						Intercept = $-0.65 \pm 0.54$
		Strain	0.71	1	0.39	
	Escape strategy	Food	1.11		0.29	
	(n = 73)	Sex	0.68	1	0.40	
		Strain × Food	4.17	2	<0.05	Farm-strain conventional =
						Intercept = $4.68 \pm 0.41$
	Exploration	Strain	0.08	1	0.78	
	(square root, n	Food	2.16	1	0.15	
	= 73)	Sex	0.89	1	0.34	
Open-field		Strain × Food	0.11	2	0.11	
test						$Intercept = 9.14 \pm 0.87$
	Activity (square	Strain	14.26	1	<0.001	Farm-strain = - 3.35 ± 0.88
	root, n = 73)	Food	3.52	1	0.06	
		Sex	0.22	1	0.90	

	Strain x Food	0.30	2	0.58	
					Intercept = $3.04 \pm 0.81$
<b>F</b>	Strain	8.19	1	<0.01	$Farm-strain = -1.80 \pm 0.70$
Escape strategy $(n = 73)$	Food	1.15		0.28	
(n - 75)	Sex	0.70	1	0.40	
	Strain $\times$ Food	1.84	2	0.17	
Duration of					Intercept = $3.17 \pm 0.20$
flight	Strain	12.85	1	<0.001	$Farm-strain = -0.94 \pm 0.26$
(for birds that	Food	7.30	1	< 0.01	$Conventional = -0.67 \pm 0.25$
flew, n = 57)	Sex	2.07	1	0.16	
	Strain $\times$ Food	5.07	2	0.03	Farm-strain conventional =

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Table 3 Summary of escape strategy (flying versus running) during cage and open-field tests with the percentage of partridges who tended to escape by flying rather than running depending on strain (wild vs. farm) and food type (organic vs. conventional grains).

	Wild strain		Farm strain	
	Organic	Conventional	Organic	Conventional
Partridges flying during cage test	39%	50%	72%	37%
Partridges flying during open- field test	89%	94%	78%	53%

620

# 621 Figures captions

Fig. 1 Schematic representation of the experimental design, involving consecutives (A)
outside FID test (B) inside open-field behavioural tests on game farm or wild grey partridges
fed organic or conventional grains.

625

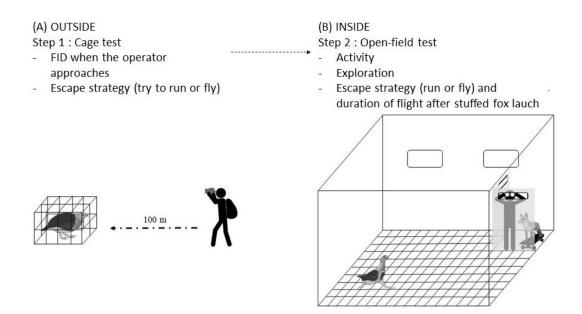
Fig. 2 Temporal variation of (a) haematocrit and (b) body mass index over time, from the 626 beginning (week 0) to the end (week 12) of the food type experiment, with different strains (wild 627 vs. farm) and food types (organic vs. conventional grains). The horizontal line corresponds to the 628 629 median, the top and bottom of boxes are the first and third quartiles, and whiskers show the lower 630 and higher values included in the 95% confidence interval. Black points correspond to the mean, open dots correspond to outliers. Stars correspond to significant effects of post-hoc tests and 631 Bonferroni correction for multiple comparisons when the interaction between food type and strain 632 was significant (i.e., p < 0.05). 633

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**Fig. 3** Variation in (a) flight initiation distance, (b) activity and (c) duration of flight according to strain (wild vs. farm) and food type (organic vs. conventional grains). The horizontal line corresponds to the median, the top and bottom of boxes are the first and third quartiles, and whiskers show the lower and higher values included in the 95% confidence interval. Black points correspond to the mean, open dots correspond to outliers.

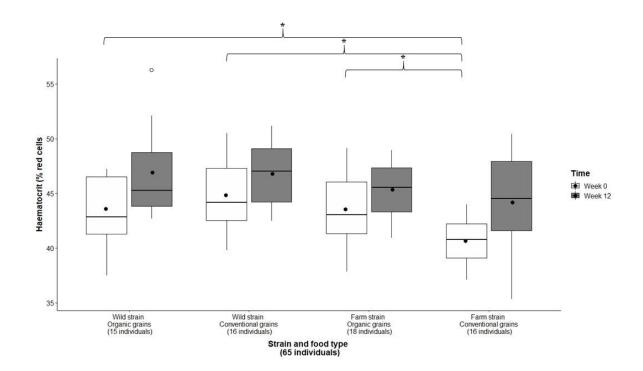
640

# 642 Fig 1

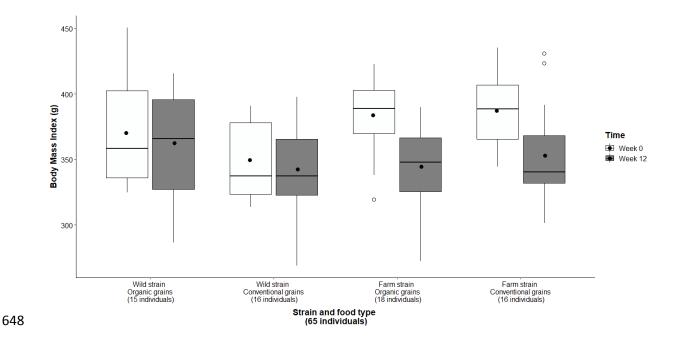


643

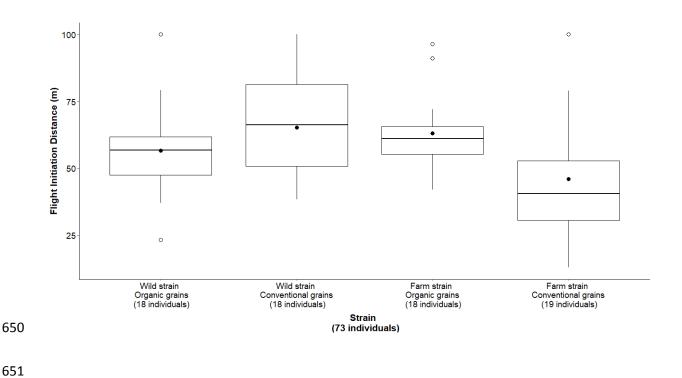
644 Fig 2.a



647 Fig 2.b



649 Fig 3.a



653 Fig 3.b

