# Distribution of Pomphorhynchus laevis s.l. (Acanthocephala) among fish species at a local scale: importance of fish biomass density 

Marie-Jeanne Perrot-Minnot, Loïc Bollache, Clément Lagrue

## - To cite this version:

Marie-Jeanne Perrot-Minnot, Loïc Bollache, Clément Lagrue. Distribution of Pomphorhynchus laevis s.l. (Acanthocephala) among fish species at a local scale: importance of fish biomass density. Journal of helminthology, 2020. hal-04480950

## HAL Id: hal-04480950

## https://u-bourgogne.hal.science/hal-04480950

Submitted on 27 Feb 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Distribution of Pomphorhynchus laevis s.l. (Acanthocephala) among fish species at a local scale: importance of fish biomassdensity 

<br>${ }^{\text {a }}$ Biogéosciences, UMR 6282 CNRS, Université Bourgogne Franche-Comté, 6 Boulevard Gabriel, 21000 Dijon, France<br>${ }^{\text {b }}$ Laboratoire Chrono-environnement, UMR 6249 CNRS, Université Bourgogne FrancheComté, 16 Route de Gray, 25000 Besançon, France<br>${ }^{c}$ Department of Biological Sciences, University of Alberta, Edmonton, T6G 2E9, Alberta, Canada

§ Both authors contributed equally to this work

Running headline: Aggregation in Pomphorhynchus spp.

* Author for correspondance

Biogéosciences, UMR 6282 CNRS, Université Bourgogne Franche-Comté
6 Boulevard Gabriel, 21000 Dijon, France



#### Abstract

Summary

Parasite distribution among hosts is a fundamental aspect of host-parasite interactions. Aggregated parasite distributions within and across host species are commonly reported and potentially influenced by many factors, whether host or parasite specific, or related to hostparasite encounter and compatibility. Yet, the respective role of each in observed parasite distributions are often unclear. Here, we documented the distribution of the acanthocephalan parasite Pomphorhynchus laevis sensu lato in two replicate fish host populations. Aggregated distributions were observed in both populations, within and across fish host species. We found positive abundance-prevalence relationship across fish species, suggesting that resource availability (fish host biomass-density) was the main driver of P. laevis s.l. distribution. This was supported by further positive associations between mean parasite load and fish biomassdensity. We found little evidence for intensity-dependent regulation within host (i.e. intra-host competition among co-infecting parasites). Furthermore, P. laevis s.l. infection had no detectable effect on fish condition indices, except on the body condition of female barbel (Barbus barbus). Pomphorhynchus laevis s.l. therefore tended to accumulate with size/age within fish species, and with fish biomass density among fish species, with apparently negligible limitations due to intra-host intensity-dependent regulation of parasite, or to parasite-induced morbidity in fish. The relative availability of final hosts for trophictransmission thus appears to be the main driver of $P$. laevis s.l. distribution among fish.


Key-words: aggregation, body-condition, Cyprinid fish, complex life-cycle, freshwater, gonadosomatic index, hepatosomatic index, overdispersion.

## Introduction

The spatio-temporal distribution of parasites among hosts is a fundamental and dynamic aspect of host-parasite interactions (Shaw \& Dobson, 1995, Poulin, 2007a; Morand \& Krasnov, 2008; Poulin et al., 2011). Heterogeneity in parasite distribution within and among host species commonly expresses as over-dispersion (or aggregation), a pattern reported in a diversity of host-parasite systems (Shaw \& Dobson, 1995; Wilson et al., 2002; Morand \& Krasnov, 2008). Aggregated parasite distribution mainly arises by chance, due to random variations associated with parasite encounter and successful establishment (Poulin, 2013; Gourbière et al., 2015). The level of intra and interspecific parasite aggregation among hosts is also expected to fluctuate with non-random variation in exposure rate and infection success (i.e. the probability of encounter and host-parasite compatibility) (Anderson \& Gordon, 1982; Shaw \& Dobson, 1995; Wilson et al., 2002; Poulin, 2007a, b; Perez-del-Olmo et al., 2011; Poulin, 2013; Gourbière et al., 2015; Johnson \& Wilber, 2017). Such heterogeneity is associated with host and parasite specific features, from individual to species level. For instance, variation in encounter probability may arise from spatio-temporal differences in foraging strategies or habitat preferences among hosts, or from differences in parasite-induced alterations of host behaviour that modulate transmission success. Variations in the rate of infection success may arise from differences in host physiological defence systems, and in parasite exploitation or evasion strategies. Parasite species-specific features, such as life cycle (direct or complex) and transmission mode (e.g. passive, active or trophic; Shaw \& Dobson 1995; Wilson et al., 2002; Poulin, 2007a), also determine parasite distribution among hosts.

Parasites with complex (i.e. heteroxenous) life cycles and trophic transmission rely on the consumption of infected intermediate or paratenic hosts by suitable definitive hosts, to complete their life cycle. They are expected to accumulate in definitive hosts, especially those occupying higher trophic levels, as these hosts are likely to consume large numbers of
infected intermediate host prey (Shaw \& Dobson, 1995; Perez-del-Olmo et al., 2011; Lester \& McVinish, 2016). At the intraspecific level, they may also accumulate with predator size and age, as prey uptake increases with predator body size. This should be especially true for parasites with a prolonged use of their host for growth and reproduction, unless intensitydependent regulation occurs.

In this context, acanthocephalan parasites offer interesting features to test these predictions (Kennedy, 2006). They have a two-host life cycle involving arthropods as intermediate hosts and vertebrates as definitive hosts, occasionally incorporating paratenic hosts (Crompton \& Nickol, 1985; Kennedy, 2006; Médoc et al., 2011). Arthropods become infected when accidentally consuming eggs. In the intermediate host, the parasite grows and then enters its last developmental stage (cystacanth), waiting for trophic transmission to the definitive host. Upon predation of the intermediate host by the appropriate vertebrate definitive host, further growth, sexual maturation and reproduction take place, and eggs are released with host faeces (Crompton \& Nickol, 1985). Acanthocephalans have a prolonged use of their vertebrate hosts for growth, sexual maturation and continuous reproduction (Crompton \& Nickol, 1985). Accumulation of intestinal parasites is thus expected to occur over time, although intensity-dependent regulation of parasite infra-population within individual hosts may limit parasite accumulation. In addition to heterogeneity in spatial distribution, abundance of adult acanthocephalans may also show marked seasonal variations (Crompton \& Nickol, 1985; Dudiňák and Špakulová, 2003; Kennedy, 2006

The goal of this study was precisely to address the distribution of the acanthocephalan parasite of freshwater fish, Pomphorhynchus laevis sensu lato Amin et al., (2003) (thereafter P. laevis s.l.), within local fish communities. Previous records of $P$. laevis s.l. suggest a broad range of fresh and brackish water fish species as definitive hosts and amphipods as intermediate hosts across the Western Palaearctic area (Kennedy, 2006; Špakulová et al.,

2011; Vardić Smrzlić et al., 2015; Perrot-Minnot et al., 2018), but also at local scales (PerrotMinnot et al., 2019). Instead of comparing distribution patterns among host-parasite systems (Morand \& Krasnov, 2008) or among populations within a given host-parasite system (Rodríguez \& Valdivia, 2017), we focused on inter-host heterogeneity in parasite distribution at a local scale. We considered the assemblage of local fish species as resource patches structuring the parasite population into infra-populations. Specifically, we assessed parasite distribution across a local range of fish species, and its consequence on fish health. We aimed at answering the following questions. (1) Are the patterns of mean abundance and aggregation of $P$. laevis s.l. within local fish host community assemblages consistent with some general macroecological laws? We predicted that mean parasite abundance should be positively correlated to prevalence (Morand \& Krasnov, 2008), that parasite abundance should vary with fish age (Anderson \& Gordon, 1982) approximated by fish size, and that heterogeneity in Pomphorhynchus distribution within fish host species should be consistent with the general pattern of parasite aggregation (Shaw \& Dobson 1995; Wilson et al., 2002; Morand \& Krasnov, 2008). (2) Is variation in mean parasite abundance among fish species related to local fish biomass-density, or density (Arneberg et al., 1998, Arneberg, 2001; Poulin, 2007a; Buck \& Lutterschmidt, 2017)? This prediction holds if encounter rate of infected intermediate host prey with a given predator species increases with predator density, with little effect of other fish-specific features such as differences in diet or compatibility towards $P$. laevis s.l. (3) Is variation in aggregation levels among fish species related to fish density? We expect a positive association between parasite aggregation levels and fish density or biomass-density across fish species, as suggested in previous studies, but also across host populations for a given host-parasite system, (Rodriguez \& Valdivia, 2017: Johnson \& Wilber, 2017). (4) Is there evidence for negative effects of infection on fish condition?

We documented the pattern of host use by $P$. laevis s.l. in two rivers, using standard infection parameters (Shaw \& Dobson, 1995; Morand \& Krasnov, 2008). We conducted standard analysis of abundance and distribution of $P$. laevis s.l. within the fish host network, using abundance - prevalence and abundance - variance relationships (Morand \& Krasnov, 2008). Abundance-occupancy relationship is a general pattern in free-living species (Gaston et al., 2000), and in parasites (Morand \& Krasnov 2008). At the intraspecific level, abundance-occupancy relationship is expected to be driven by temporal variations in resource availability (Gaston et al., 2000). For a given parasite species, variations in resource availability may arise within the community of host species (Morand \& Krasnov 2008), driven by relative host density or biomass-density. We estimated P. laevis s.l. aggregation among fish host species using the abundance-variance relationship known as Taylor's power law (Anderson \& Gordon, 1982; Shaw \& Dobson, 1995; Kilpatrick \& Ives, 2003; Morand \& Krasnov, 2008; Johnson \& Hoverman, 2014). Finally, we assessed potential effects of $P$. laevis s.l. infection on fish health, by estimating three standard body-condition metrics: body condition index (BCI), hepatosomatic index (HSI, related to energy storage), and gonadosomatic index (GSI, reflecting reproductive investment) (Cheppala et al., 1995). These metrics are commonly used to assess the impact of pollutants (Dragun et al., 2013), infection (Tierney et al., 1996; Kalogianni et al., 2013; Masson et al., 2015), and habitat quality (Nagrodski et al., 2013) in fish.

## Material and methods

## Localities, fish community composition and sampling

One locality on the Ouche River ( $47^{\circ} 17^{\prime} 54.56{ }^{\prime \prime} \mathrm{N} 5^{\circ} 2^{\prime} 21.97{ }^{\prime \prime} \mathrm{E}$ ), and one locality on the Vingeanne River ( $47^{\circ} 20^{\prime} 51.66^{\prime \prime} \mathrm{N} 5^{\circ} 27^{\prime} 8.76$ "E) were sampled in 2003 and 2005, and in 2004
and 2005, respectively. To avoid incorporating seasonal variation in prevalence and abundance to the analysis of $P$. laevis s.l. distribution, we collected samples at the same time of year (late spring / early summer). We retrieved information on the composition of local fish communities in these two localities from the ONEMA database (http://www.naiades.eaufrance.fr/acces-donnees\#/hydrobiologie), based on the regular monitoring of fish species richness and abundance between 2001 and 2006. The database provides a full record of density and biomass-density (thereafter biomass) of each fish species locally present. Thirty fish species were identified of which fourteen were present in both localities (Fig. S1 in Perrot-Minnot et al., 2019).

Fish were captured by electric fishing or netting, killed immediately, identified, measured (fork length) and weighed. Upon fish dissection, we removed, opened and screened intestines to collect adult $P$. laevis s.l. Since the occurrence of paratenic hosts or dead-end hosts has been previously reported for P. laevis s.l. (Crompton \& Nickol, 1985), we also inspected fish body cavity and viscera for extra-intestinal infection with P. laevis s.l. cystacanths. We included in the analyses only fish species for which at least five individuals were dissected. We measured gonad and liver weights on a subset of fish to estimate GSI and HSI.

We did not distinguish $P$. tereticollis and $P$. laevis in the present study, despite the recent taxonomic revision of $P$. laevis s.l. and the erection of $P$. tereticollis as a true species (Špakulová et al., 2011). Both species are present in the two localities sampled here, but they share the same amphipod intermediate hosts and fish definitive host species. Although they seem to exhibit some specificity towards fish final hosts in terms of relative abundance rather than presence/absence (Perrot-Minnot et al . 2019), a preliminary analysis suggested that both Pomphorhynchus species exhibited comparable level of aggregation among fish host. This subset of 815 Pomphorhynchus spp. required genotyping for identification, and was not large
enough to quantify reliably parasite distribution pattern among all fish species for each Pomporhynchus species. In fact, 2005 Pomphorhynchus that were not genotyped, and 268 additional individual fish (including non-infected ones), were added to the dataset used to run the present analysis.

Infection pattern of P . laevis s.l.: prevalence, abundance and aggregation across fish host species

We used multiple descriptors to characterize $P$. laevis s.l. infection patterns in each fish species (Bush et al., 1997): prevalence (i.e. proportion of individual hosts infected), abundance (i.e. mean number of parasites per host), and intensity (i.e. mean number of parasites per infected host). We estimated Pomphorhynchus laevis s.l. prevalence and intensity for two non-exclusive categories of individual host: fish infected with intestinal adult P. laevis s.l. and fish harbouring cystacanths in the body cavity and in viscera. We performed the analysis of $P$. laevis s.l. abundance and aggregation only on intestinal adult parasites, since we considered the distribution pattern of parasites among hosts in relation to population stability. Indeed, the contribution of extra-intestinal parasites to the parasite life-cycle and population dynamics is unclear, as it may depend on the host species considered (i.e. some may act as paratenic hosts, given their inclusion into the diet of piscivorous fish, while others are dead-end hosts, such as minnow and catfish, respectively) (Médoc et al., 2011; PerrotMinnot et al., 2019).

We computed the distribution of $P$. laevis abundance for each fish species within each river, to illustrate its distribution pattern among individual fish for each fish species. We tested the relationship between $P$. laevis s.l. abundance ( $\log 10+1$ transformation) and individual body size using linear regression.

We estimated the degree of aggregation of intestinal adult P. laevis s.l. within fish species using the slope of the regression of variance in parasite abundance to mean parasite abundance (both $\log 10$-tranformed), known as Taylor's power law, among fish species. To get close to the sample size recommended (> 30; Shaw and Dobson 1995; Poulin 2013), we removed fish species for which less than 25 individuals were sampled, and run the analysis on data from the Ouche River only ( $\mathrm{N}=9$ species of fish). Departure from random distribution of parasites towards aggregation among fish hosts would be evidenced by a slope greater than one (Morand \& Krasnov, 2008; Johnson \& Hoverman, 2014).

The variance-to-mean ratio was then used as a measure of aggregation level, in preference to the other commonly used parameter k (Morand \& Krasnov, 2008), because we compared parasite distribution patterns among fish species differing in P. laevis s.l. prevalence (and for some of them, with a large number of uninfected hosts) (Scott, 1987 in Wilson et al., 2002). We used the variance-to-mean ratio to test for a relationship between aggregation level and fish biomass, across the range of fish species.

## Fish condition in relationship to P. laevis s.l. abundance

Variations in fish (BCI, HSI and GSI were analysed in relation to intestinal P. laevis s.l. abundance, fish species and sex. Only fish species and sex for which at least twelve individuals were screened and two infected were included in the analyses. Some individual fish could not be sexed and were included as "juveniles". We measured fish body condition index as the residuals of the regression of $\log 10$-transformed whole body mass on $\log 10$ transformed body size (fork length) (Chellappa et al., 1995). Since the relationship was significantly different among fish species, the residuals were estimated for each fish species separately. Hepatosomatic and gonadosomatic indices were calculated as the ratio of liver weight and gonad weight on body mass, respectively (Chellappa et al., 1995)

## Data analysis

All statistical analyses were performed using $R$ software v 3.5.1. and v.3.6.1. (R Core team, 2018). Within each fish species, the relationship between P. laevis s.l. abundance ( $\log _{10}+1$ transformation) and individual body size was tested using linear regression. We also computed the $95 \%$ confidence intervals around prevalence using PropCIs package (v. 0.3.0) (Scherer, 2018).

We run linear and generalized linear models (GLM) to analyse prevalence and abundance, We performed model comparison to estimate the contribution of each predictor variable to variation in the dependant variable. The approach is based on deviance comparison between models fitted to the same data - the full model and the model without one predictor variable- (maximum likelihood ratio test, Rpackage 'lmtest', v. 0.9-36; Zeileis \& Hothorn, 2002). We used the associated Chi-square value and probability to assess the significance of each predictor variable. We also reported the coefficient of determination $\mathrm{R}^{2}$, as an estimate of how well variation in the dependant variable is explained by predictor ones. As we included locality as a random factor, we computed both the marginal $\mathrm{R}^{2} \mathrm{~m}$ and the conditional $\mathrm{R}^{2} \mathrm{c}$, which represents the part of the variance explained by fixed effects and by the entire model respectively, using R package 'MuMIn' v. 1.42.1 (Barton, 2018). We analysed differences in prevalence and abundance according to fish species using GLM with binomial distribution, and GLM with negative binomial link function, respectively, adding locality as a random factor (package Lme4 v.1.1.19, Bates et al. 2015; and package MASS v.7.3.51; Venables \& Ripley 2002, respectively). We compared parasite intensity among fish species within each locality using a Kruskal-Wallis test and post-hoc paired comparisons (Dunn test, Benjamini Yekutieli B-Y method of correction for multiple tests; package Dunn-test, Dinno, 2015) as the distribution of intensity could not be normalized.

We analysed the relationship between prevalence and mean abundance across the range of fish species using spearman rank correlation test. We tested the relationship between mean P. laevis s.l. abundance and fish density or biomass across the range of fish species using linear regression after $\log -\log$ transformation ( $\log 10$ ), adding locality as a random factor.

We analysed the aggregation of $P$. laevis s.l. across the nine fish species from Ouche River for which more than 25 individuals were sampled. We regressed the $\log 10$-transformed variance of abundance on the $\log 10$-transformed mean abundance, applying a simple linear model, Given the low sample size ( 9 fish species), we used ordinal nonparametric boostrap procedure (1000 replicates) to get the bootstrapped value of the regression slope and its $95 \%$ confidence interval (Package boot, v.1.3-23) (Canty \& Ripley, 2019). We then tested for an effect of fish density- biomass on the variance-to-mean abundance ratio, using the Spearman rank correlation test.

All three condition parameters, BCI, HIS and GSI were used as the dependent factor in linear model (BCI) or GLM with gamma-distribution (HIS, GSI), including parasite abundance ( $\log 10$-transformed), sex, and their interaction, as fixed effects. Fish species and its interaction with parasite abundance were included in the full model for HIS and GSI, while separate analyses of BCI were done for each fish species.

## Results

Pomphorhynchus laevis s.l. samples were collected from 14 species of fish, mainly cyprinids, among which four species were collected in both localities (Table 1). A total of 881 fish were dissected; 752 from 11 fish species sampled in the Ouche River and 129 from 9 fish species sampled in the Vingeanne River. The sampling of each fish species was representative of the
local fish community, irrespective of whether the fish species hosted P. laevis s.l. or not (see Table S1 in Perrot-Minnot et al., 2019).

Prevalence and abundance of P. laevis s.l.
Intestinal P. laevis s.l. were collected from all of the eleven fish species sampled in the Ouche River and from six out of the seven fish species in the Vingeanne River (Table 1; Fig. 1). Extra-intestinal P. laevis s.l. cystacanths were found in several species of fish; all but one (Chondrostoma nasus) also harboured intestinal adults (Table 1; Fig. 1). Extra-intestinal parasites were either embedded as cystacanth or attached as evaginated cystacanth on liver and gonad surface or in adipose tissues.

Parasite distribution within each fish species was clearly over-dispersed (Fig. S1). At the intra-host level, P. laevis s.l. abundance (log-transformed) increased significantly with individual fish size in most fish species in both localities, but with a low to medium coefficient of determination ( 0.07 to 0.62 ) (Fig. S2). There was no evidence for a decrease in mean and variance of abundance in the upper-size category, but rather a log-linear trend for parasite load to increase with body size (Fig. S2).

Prevalence of intestinal P. laevis s.l. differed among fish species $\left(\mathrm{Chi}^{2}=171.3, \mathrm{df}=\right.$ $12, P<0.0001, \mathrm{R}^{2} \mathrm{~m}=0.21, \mathrm{R}^{2} \mathrm{c}=0.37$ ), as well as both intestinal and extra-intestinal $P$. laevis s.l. $\left(\mathrm{Chi}^{2}=117.9, \mathrm{df}=13, P<0.0001, \mathrm{R}^{2} \mathrm{~m}=0.34, \mathrm{R}^{2} \mathrm{c}=0.49\right)$. Half of fish species harboured prevalence above $50 \%$ (Fig. 1). Infection intensity of intestinal P. laevis s.l. differed among fish species in the Ouche River (Kruskal-Wallis chi-squared $=43.1724, \mathrm{df}=$ $10, P<0.0001$ ) but not in the Vingeanne River (chi-squared $=2.44, \mathrm{df}=5, P=0.79$ ) (Fig. S3). There was no effect of sampling effort (number of individual fish sampled per fish species) on prevalence or intensity (Spearman correlation test, samples from both rivers pooled: $\mathrm{N}=18 ; \mathrm{S}=1254, P=0.24$, $\mathrm{Rho}=-0.29$; and $\mathrm{S}=11.66, P=0.15$, $\mathrm{Rho}=0.66$,
respectively). The abundance of intestinal $P$. laevis s.l. differed among fish species $\left(\mathrm{Chi}^{2}=\right.$ 436.4, df $=12, \mathrm{P}<0.0001, \mathrm{R}^{2} \mathrm{~m}=0.38, \mathrm{R}^{2} \mathrm{c}=0.54$ ). Abundance and prevalence were significantly correlated across fish species and localities (Spearman correlation test: $\mathrm{N}=17, \mathrm{~S}$ $=167.6, P=0.0001$, Rho $=0.79$; Fig. 2).

## Are differences in mean abundance among fish hosts explained by local fish biomass?

We tested the hypothesis that mean parasite abundance per fish species could be determined by host species abundance. The mean abundance of $P$. laevis s.l. in fish species was positively correlated with local fish biomass $\left(\mathrm{N}=17 ; \mathrm{Chi}^{2}=4.76, \mathrm{df}=1, P=0.03 ; \mathrm{R}^{2} \mathrm{~m}\right.$ $\left.=\mathrm{R}^{2} \mathrm{c}=0.25\right)\left(\right.$ Fig. 3), but not with local fish density $\left(\mathrm{Chi}^{2}=1.53\right.$, $\mathrm{df}=1, P=0.22$ (Fig. S4). The roach Rutilus rutilus, stood as an outlier in the Ouche River but not the Vingeanne River, with a lower mean abundance than expected from its biomass (Fig. 3).

Aggregation of intestinal P. laevis s.l. across fish species
The level of aggregation of intestinal P. laevis s.l. among fish species was estimated using the relationship between variance in parasite abundance and mean abundance (log 10transformed) among nine fish species from the Ouche River. Variance in abundance of intestinal parasites among fish species was strongly related to mean abundance, with a slope significantly different from unity, indicative of aggregation relative to random distribution (Table 2, Fig. 4). The mean abundance of intestinal P. laevis s.l. explained more than $95 \%$ of the variance in abundance. Ouche RiverVingeanne RiverOuche RiverVingeanne River

The variance-to-mean abundance ratio tended to increase with biomass-density although not significantly (bootstrapped estimate of Spearman rank correlation and its 95\% CI: $\mathrm{N}=9$, Rho $=0.68[-0.34-0.97])($ (Fig. S5 $)$.

## Effect of infection on fish condition indices

Variations in BCI were not explained by the full model including parasite abundance, host sex and their interaction as predictors in any of the fish species, except in barbel from Ouche River (Table 3a), where body condition decreased with parasite load in females (Fig. S6).

The relationship between hepatosomatic index and parasite load differed among fish species (Table 3b). Interestingly, HIS increased significantly with parasite load in barbel (Fig. S7a); however, HIS increased significantly with body size in the same manner (Fig. S7b). Variations in gonadosomatic index were independent of parasite load (Table 3b). GSI differed according to fish species and sex (Fig. S7c, d), with females exhibiting higher size-corrected weight of gonads compared to males and juveniles (Fig. S7d).

## Discussion

## Abundance-prevalence relationship and aggregation

The distribution of $P$. laevis s.l. within most fish species was aggregated, confirming the pattern reported in other Acanthocephalan species, both in final hosts (Anderson \& Gordon, 1982; Dobson \& Keymer, 1985; Kennedy 2006), and in intermediate hosts (Dobson \& Keymer, 1985; Rodríguez \& Valdivia, 2017), and more generally in macroparasites (Shaw \& Dobson, 1995; Poulin, 2007a, Morand \& Krasnov, 2008).

The more abundant $P$. laevis s.l. was in a given host species, the highest was its prevalence, in both localities. This pattern is equivalent to the abundance-occupancy relationship in ecology. Variations in habitat availability and quality can generate such pattern in free-living species (Gaston et al., 2000, Freckleton et al., 2006). For parasites, it corresponds to variation in host availability (encounter and transmission rates) and compatibility. To further understand parasite distribution across fish species, we analysed
aggregation using the variance - to - mean abundance relationship (Gaston et al., 2000; Morand \& Krasnov, 2008), on a subset of fish species. A high proportion of variance in parasite abundance (95\%) was explained by mean abundance across fish species. This result is in agreement with Shaw \& Dobson (1995) and Poulin (2013), and provides evidence for a nearly random process of $P$. laevis s.l. accumulation among fish species. Several factors could potentially increase heterogeneity in parasite distribution among fish host species, such as sample size and host mean body size (Poulin, 2013; Johnson \& Wilber, 2017), and behavioural or physiological differences among host species (Shaw \& Dobson 1995). Differences in diet choice, microhabitat and host-parasite compatibility can all influence parasite distribution among hosts. However, their contribution to variation in aggregation among fish hosts should be negligible compared to the process of random accumulation of $P$. laevis s.l. This conclusion should be taken cautiously however; sample size limitations restricted the number of fish species included to only nine out of the fourteen species of fish hosts recorded.

## Abundance and aggregation in relation to fish biomass

The process of random accumulation of P. laevis s.l. evidenced here was suggesting that parasite distribution within and among fish species was mainly driven by the probability of encounter rate of predators with infected host prey. We attempted to test this hypothesis by using local fish biomass-density as a proxy for encounter rate. Interestingly, we found a positive relationship between mean parasite abundance and fish biomass-density, but not fish density. Most fish species prey upon amphipods, and these crustaceans are the most abundant macroinvertebrates sampled in these rivers (MJPM pers. observation), Therefore, encounter rate with infected prey increases with fish host density but also body mass, both leading to a higher prey intake rate. The distribution of $P$. laevis s.l. among fish hosts further confirms that
potential variations in compatibility across fish host species have a negligible effect on the distribution of $P$. laevis s.l. This interpretation holds assuming that the accumulation of adult intestinal parasites in individual fish occurs at a rate proportional to encounter rate across fish species, meaning low heterogeneity in compatibility, and low within-host competition among co-infecting parasites. In support of this assumption, the lack of intensity-dependent regulation of parasite development or fecundity has been reported previously on a subset of these samples. Parasite load had no effect on parasite body size or reproductive parameters (testes volume and number of eggs) (Perrot-Minnot et al., 2019). It is still possible that interindividual differences within fish species in growth rate or diet choice contribute to the low to medium coefficient of determination between parasite load and fish size. In addition, the occurrence of extra-intestinal parasites in several fish species still suggests that some species are more suitable definitive hosts than others. However, the analysis of aggregation pattern using all P. laevis s.l. (both intra- and extra-intestinal) points to the same conclusions (data not shown). We favoured the analysis of intestinal parasite distribution, since the contribution of extra-intestinal parasites to the life-cycle is difficult to establish. Indeed, it may differ according to fish host, either paratenic hosts (minnow), or dead-end hosts (catfish), the former being part of piscivorous fish diet.

Given the relation of parasite distribution to local fish biomass density, a large fraction of $P$. laevis s.l. population typically occurs within a few host species, mainly barbel and chub. Interestingly, this pattern does not preclude narrower host range when considering $P$. laevis and $P$. tereticollis separately, in particular with respect to fish feeding ecology (Perrot-Minnot et al., 2019).

## Effects on fish host condition/health

We did not observe a decrease in the mean and variance of abundance in the upper category of fish size. Following Lester (1982) and Wilson et al. (2002), and assuming that size increases with age, this log-linear increase in abundance with age suggests negligible parasiteinduced mortality or acquired immunity in fish. In addition, no effect of infection on body condition was evident, except in female barbel, that exhibited a decreasing body condition with increasing parasite load. Since barbel exhibited the highest mean intensity of $P$. laevis s.l., with $25 \%$ of individuals harbouring more than 30 adult worms (Table 1; Fig. S1), it is possible that $P$. laevis s.l. impacts fish body condition when reaching intensities hardly seen in most fish species. In addition, parasite load did not affect HIS nor GSI. The spurious positive correlation between HIS and parasite load found in barbel was likely the consequence of the positive correlations of both HIS and parasite load with fish body size (or age). As such positive relationship between HIS and host size has been reported before (in sharks, Hussey et al., 2009), it is important to emphasize the necessity to look for confounding variables before interpreting the relationship between HIS and parasite load. In ectotherms with continuous growth, this relationship is probably reflecting both the higher metabolic efficiency of larger individuals within a predatory species, possibly sustained by a larger liver relative to body mass or size, and the increase in parasite load with age (approximated by size). To our knowledge, this hypothesis has never been addressed so far.

## Conclusion

Positive relationships between abundance and occupancy, and variance and mean in abundance have been regularly evidenced at the intraspecific and interspecific levels, in freeliving (Gaston et al., 2000) and parasite species (Morand \& Krasnov, 2008; Jenkins \& Owens, 2011; Pérez-del-Olmo et al., 2011). Yet, the underlying causes are not well understood, as they potentially include diverse intrinsic and extrinsic factors (reviewed in Johnson \&

Hoverman 2014, Poulin 2007a). Here, the positive association between mean P. laevis s.l. abundance and fish biomass-density tends to suggest that resource availability, represented by fish biomass-density, is the main driver of parasite distribution among hosts. It is important to notice that such relationship did not hold using fish density. Pomphorhynchus laevis s.l. therefore tends to accumulate with age within fish species, and with fish biomass-density among fish species, with negligible limitations due to intra-host intensity dependent regulation of parasite nor of parasite-induced morbidity in fish, respectively. It shows that biomass-density, rather than density alone, has to be taken into account for trophically transmitted parasites, by contrast to parasites transmitted by contact. We thereby emphasized a specific feature of spatial distribution of 'predators as resources' for trophically-transmitted parasites; and its contribution to aggregative distribution.

## Acknowledgments

We thank the Agence française pour la biodiversité (Mr Julien BOUCHARD) for his assistance during to the field monitoring campaign. Experiments and fish sampling comply with the current laws of France. This work was supported in part by a grant from the Biologial Invasion program no. 01121 (Ministère de l'Environnement et du Développement durable) and by a grant (contrat d'étude) from the Conseil Régional de Bourgogne.

## References

Amin, O.M., Abdullah, S.M.A. \& Mhaisen, F.T. (2003) Description of Pomphorhynchus spindletruncatus n . sp (Acanthocephala : Pomphorhynchidae) from freshwater fishes in northern Iraq, with the erection of a new pomphorhynchid genus, Pyriproboscis n. g., and keys to genera of the Pomphorhynchidae and the species of Pomphorhynchus Monticelli, 1905. Systematic Parasitology 54, 229-235.

Anderson, R.M. \& Gordon, D.M. (1982) Processes influencing the distribution of parasite numbers within host populations with special emphasis on parasite-induced host mortalities Parasitology 85, 373-398.

Arneberg, P. (2001) An ecological law and its macroecological consequences as revealed by studies of relationships between host densities and parasite prevalence. Ecography 24, 352-358.

Arneberg, P., Skorping, A., Grenfell, B. \& Read, A.F. (1998) Host densities as determinants of abundance in parasite communities. Proceedings of the Royal Society B: Biological Sciences 265, 1283-1289.

Barton, K. (2018) MuMIn: Multi-Model Inference. R package version 1.42.1.
Bates, D., Maechler, M., Bolker, B. \& Walker, S. (2015) Fitting Linear Mixed-Effects Models Using lme4. Journal of Statistical Software 67, 1-48. doi:10.18637/jss.v067.i01.

Buck, J.C. \& Lutterschmidt, W.I. (2017) Parasite abundance decreases with host density: evidence of the encounter-dilution effect for a parasite with a complex life cycle. Hydrobiologia 784, 201-210.

Bush, A.O., Lootvoet, K.D., Lotz, J.M. \& Shostak, A.W. (1997) Parasitology meets ecology on its own terms: Margolis et al. revisited. Journal of Parasitology 83, 575-583.

Canty, A. \&Ripley, B. (2019). boot: Bootstrap R (S-Plus) Functions. R package version 1.323.

Chellappa, S., Huntingford, F.A, Strang, R.H.C., \&Thomson, R.Y. (1995) Condition factor and hepatosomatic index as estimates of energy statusin male 3-spined stickleback. Journal of Fish Biology 47, 775-787.

Crompton, D. W. T. \& Nickol, B. B. (1985) Biology of Acanthocephala. 1st edn. 519 pp. Cambridge, Cambridge University Press.

Dinno, A. (2015) Dunn.test: Dunn's Test of Multiple Comparisons Using Rank Sums. R package version 1.3.1.

Dudiňák, V. \& Špakulová, M. 2003. The life cycle and seasonal changes in the occurrence of Pomphorhynchus laevis (Palaeacanthocephala, Pomphorhynchidae) in a small isolated lake. Parasite 10, 257-262.

Dragun, Z., Filipović Marijić, V., Kapetanović, D., Valić, D., Vardić Smrzlić, I., Krasnići, N., Strižak, Ž., Kurtović, B., Teskeredžić E. \& Raspor, B. (2013) Assessment of general condition of fish inhabiting a moderately contaminated aquatic environment. Environmental Science and Pollution Research 20, 4954-4968.

Fredensborg, B. L. (2014) Predictors of Host Specificity among Behavior-Manipulating Parasites. Integrative and Comparative Biology 54, 149-158.

Gaston, K.J., Blackburn, T., Greenwood, J.J.D., Gregory, R.D., Quinn, R.M. \& Lawton, J.H. (2000) Abundance-occupancy relationships. Journal of Applied Ecology 37, 39-59. Gourbière, S., Morand, S. \& Waxman, D. (2015) Fundamental Factors Determining the Nature of Parasite Aggregation in Hosts. PLoS ONE 10(2): 00116893. doi:10.1371/journal.pone. 0116893 .

Hussey, N. E., Cocks, D.T., Dudley, S.F.J., McCarthy, I.D. \& Wintner, S.P. (2003) The condition conundrum: application of multiple condition indices to the dusky shark Carcharhinus obscurus. Marine Ecology Progress Series 380, 199-212.

Jenkins, T. \& Owens, I.P. (2011) Biogeography of avian blood parasites (Leucocytozoon spp.) in two resident hosts across Europe: phylogeographic structuring or the abundanceoccupancy relationship? Molecular Ecology 20, 3910-3920.

Johnson, P.T. J. \& Hoverman J.T. (2014) Heterogeneous hosts: how variation in host size, behaviour and immunity affects parasite aggregation. Journal of Animal Ecology 83, 1103-1112.

Johnson, P.T.J. \& Wilber, M.Q. (2017) Biological and statistical processes jointly drive population aggregation: using host-parasite interactions to understand Taylor's power law. Proceedings of the Royal Society B: Biological Sciences 284, 20171388. http://dx.doi.org/10.1098/rspb.2017.1388

Kalogianni, E., Kmentová, N., Harris, E., Zimmerman, B., Giakoumi, S., Chatzinikolaou, Y. \& Vanhove, M.P.M. (2017) Occurrence and effect of trematode metacercariae in two endangered killifishes from Greece. Parasitoly. Research 116, 3007-3018.

Kennedy, C.R. (2006) Ecology of the Acanthocephala. 1rst edn. 260 pp. Cambridge, Cambridge University Press.

Kilpatrick, A.M. \& Ives, A.R. (2003) Species interactions can explain Taylor's power law for ecological time series. Nature 422, 65-68.

Lester, R.J.G. \& McVinish, R. (2016) Does moving up a food chain increase aggregation in parasites? Journal of The Royal Society Interface 13: pii: 20160102.

Masson, G., Vanacker, M., Fox, M.G. \& Beisel, J.N. (2015) Impact of the cestode Triaenophorus nodulosus on the exotic Lepomis gibbosus and the autochthonous Perca fluviatilis. Parasitology 142, 745-755.

Médoc, V., Rigaud, T., Motreuil, S., Perrot-Minnot, M.-J. \& Bollache, L. (2011) Paratenic hosts as regular transmission route in the acanthocephalan Pomphorhynchus laevis: potential implications for food webs. Naturwissenschaften 98, 825-835.

Morand, S. \& Krasnov, B. (2008) Why apply ecological laws to epidemiology? Trends in Parasitology 24, 304-309.

Nagrodski, A., Suski, C.D. \& Cooke, S.J. (2013) Health, condition, and survival of creek chub (Semotilus atromaculatus) across a gradient of stream habitat quality following an Pérez-del-Olmo, A., Morand, S., Raga, J. A., \& Kostadinova, A. (2011). Abundancevariance and abundance-occupancy relationships in a marine host-parasite system: the importance of taxonomy and ecology of transmission. International Journal for Parasitology 41, 1361-1370.

Perrot-Minnot, M.-J., Špakulová, M., Wattier, R., Kotlík, P., Düşen, S., Aydoğdu, A. \& Tougard, C. (2018) Contrasting phylogeography of two Western Palaearctic fish parasites despite similar life cycles. Journal of Biogeography 45, 101-115.

Perrot-Minnot, M.J., Guyonnet, E., Bollache, L. \& Lagrue, C. (2019) Differential patterns of definitive host use by two fish acanthocephalans occurring in sympatry: Pomphorhynchus laevis and Pomphorhynchus tereticollis. International Journal for Parasitology: Parasites and Wildlife 8, 135-144.

Poulin, R. (2007a) Evolutionary Ecology of Parasites. $2^{\text {nd }}$ edn. 332 pp. Princeton, Princeton University Press, USA.

Poulin R. (2007b) Are there general laws in parasite ecology? Parasitology 134, 763-776.

Poulin, R. (2013) Explaining variability in parasite aggregation levels among host samples. Parasitology 140, 541-546.

Poulin, R., Krasnov, B. R. \& Mouillot, D. (2011) Host specificity in phylogenetic and geographic space. Trends in Parasitology 27, 355-361.

R Core Team (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.Rproject.org/.computing. 2.15.0 edition. Vienna: R Foundation for Statistical Computing.

Rodríguez, S. M. \& Valdivia, N. (2017). Mesoscale spatiotemporal variability in a complex host-parasite system influenced by intermediate host body size. Peer Journal DOI 10.7717/peerj. 3675 .

## Sánchez, C.A., Becker, D.J., Teitelbaum, C.S., Barriga, P., Brown, L.M., Majewska,

 A.A., Hall, R.J. \& Altizer S. (2018) On the relationship between body condition and parasite infection in wildlife: a review and meta-analysis. Ecology Letters 21, 1869-1884.Scherer, R. (2018). PropCIs: Various confidence interval methods for proportions. R package version 0.3-0.

Shaw, D.J. \& Dobson, A.P. (1995) Patterns of macroparasite abundance and aggregation in wildlife populations: a quantitative review. Parasitology 111, S 111-S 133.

Špakulová, M., Perrot-Minnot. M.-J. \& Neuhaus, B. (2011) Resurrection of Pomphorhynchus tereticollis (Rudolphi, 1809) (Acanthocephala: Pomphorhynchidae) based on new morphological and molecular data. Helminthologia 48, 268-277.

Tierney, J.F. Huntingford, F.A. \& Crompton, D.W.T. (1996) Body condition and reproductive status in sticklebacks exposed to a single wave of Schistocephalus solidus infection. Journal of Fish Biology 49, 483-493.

Vardić Smrzlić, I., Valić, D., Kapetanović, D., Filipović Marijić, V. Gjurčević, E. \& Teskeredžić, E. (2015) Pomphorhynchus laevis (Acanthocephala) from the Sava River
basin: New insights into strain formation, mtDNA-like sequences and dynamics of infection. Parasitology Intrnational 64, 243-250.

Venables, W. N. \& Ripley, B. D. (2002) Modern applied statistics with S. Fourth Edition. Springer, New York.

Wilson, K., Bjørnstad, O.N., Dobson, A.P., Merler, S., Poglayen, G., Randolph, S., Read A.F. \& Skorping, A. (2002) Heterogeneities in macroparasite infections: patterns and processes. pp. 6-44 in Hudson P.J., Rizzoli, A., Grenfell, B.T., Heesterbeek, H. \& Dobson, A.P. (Eds) The Ecology of Wildlife Diseases Oxford University Press, Oxford, UK.

Zeileis, A. \& Hothorn, T. (2002) Diagnostic Checking in Regression Relationships. R news, 2: 7-12.

## Figure legends

Fig. 1: Prevalence of $P$. laevis s.l. in 14 fish species from two rivers. Individual fish were harbouring adult parasites in the intestine exclusively (plain bars), both adult parasites in the intestine and immature parasites (cystacanths) in body cavity or viscera (dotted bars), or exclusively immature parasites (cystacanths) in body cavity or viscera (dashed bars). Total prevalence is represented with its $95 \%$ confidence interval. (a), Ouche River; (b), Vingeanne River; $\mathrm{N}=$ number of fish sampled. Asterisk: significantly lower prevalence of intestinal parasites compared to the other groups of fish species. See Table 1 for abbreviations of fish species.

Fig. 2: Relationship between mean abundance and prevalenceof intestinal Pomphorhynchus laevis s.l. across fish host species in the Ouche and Vingeanne Rivers. This relationship corresponds to the "abundance-occupancy" rule for free-living species.

Fig. 3: Relationship between local fish biomass (g. $100 \mathrm{~m}-2$ ) and mean abundance of intestinal Pomphorhynchus laevis s.l. per fish species, across fish species and localities (Ouche and Vingeanne rivers).

Fig. 4: Relationship between variance in abundance and mean abundance of intestinal Pomphorhynchus laevis s.l. across fish host species in the Ouche River, on a log scale. Regression analysis was performed using log-transformation. A slope higher than one indicates aggregation.

Table 1 - Abundance and intensity of Pomphorhynchus laevis s.l. parasites in Ouche (O; years 2003, 2005) and Vingeanne (V; years 2004 and 2005) rivers, in fish species for which at least five individuals were sampled (loach excluded for Vingeanne River). Abundance and intensity were estimated separately for adult parasites in the intestinal track and for immature parasites (cystacanths) found outside of the intestinal track, in viscera or in the body cavity. * Prevalence of intestinal P. laevis s.l., and its $95 \%$ CI. BF: benthic fish, bottom feeder; BPF: bentho-pelagic fish.

| Fish host (habitat) | River | Number of fish sampled / infected | Number of fish with P. laevis s.l. |  | Prevalence of intestinal <br> P. laevis s.l. ( $95 \%$ CI) | P. laevis s.l. abundance (median-1 ${ }^{\text {rst }}$ and 3' quartiles) |  | P. laevis s.l.s intensity (median-1' and 3' quartiles) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | in intestinal tract | outside intestinal tract |  | Parasites in intestinal tract | Parasites outside intestinal tract | Parasites in intestinal tract | Parasites outside intestinal tract |
| All fish species | O | 752 / 377 | 375 | 54 |  | 0 (0-2) | $0(0-0)$ | $2(1-4)$ | $2(2-6)$ |
|  | V | 122 / 100 | 83 | 20 |  | $2(0-7)$ | $0(0-0)$ | $5(2-10.25)$ | $2.5(2-16.75)$ |

Cypriniformes -
Cyprinidae

| Barbus barbus (BF) | O | 26/16 | 16 | 0 | 0.61 (0.42-0.79) | $2(0-9)$ | $0(0-0)$ | 6 (2.3-31.8) | - |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Barbel, Bbs | V | $31 / 29$ | 29 | 0 | 0.94 (0.80-0.99) | $4(3-29)$ | 0 (0-0) | 5 (3-32.5) | - |
| Gobio gobio (BF) | O | 64 / 23 | 10 | 15 | 0.16 (0.08-0.26) | $0(0-0)$ | $0(0-0)$ | $1(1-2.25)$ | $2(2-6)$ |
| Gudgeon, Gg | V | $8 / 3$ | 3 | 0 | 0.38 (0.11-0.72) | $0(0-3.25)$ | $0(0-0)$ | $4(1-4)$ | . |
| Squalius cephalus (BPF) | O | 318 / 202 | 202 | 0 | 0.64 (0.58-0.69) | $1(0-3)$ | 0 (0-0) | $2(1-4)$ | - |
| Chub, Sc | V | 43 / 42 | 40 | 3 | 0.93 (0.82-0.98) | $5(2-10)$ | $0(0-0)$ | 5.5 (2-10) | $2(1-38)$ |
| Rutilus rutilus (BPF) | O | 26/1 | 1 | 0 | 0.04 (0.002-0.18) | $0(0-0)$ | $0(0-0)$ | 1 | 2 |
| Roach, Rr | V | 12 / 7 | 3 | 5 |  | 0 (0-1.5) | $0(0-2)$ | 4(2-18) | $2(1.5-9)$ |
| Telestes souffia (BPF) <br> Vairone, Ts | O | 47 / 24 | 24 | 2 | 0.51 (0.37-0.65) | $1(0-4)$ | $0(0-0)$ | $4(2-10)$ | 3,5 (1-6) |


| Chondrostoma nasus (BPF) Nase, Cn | V | 15/7 | 0 | 7 |  | $0(0-0)$ | $0(0-4)$ | - | $4(2-18)$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Table I (continued) |  |  |  |  |  |  |  |  |  |
| Fish host (habitat) | River | Number of fish sampled / infected | Number of fish with P. laevis s.l. |  | Prevalence of intestinal <br> P. laevis s.l. (95\% CI) | P. laevis s.l. abundance (median- $1^{\text {rst }}$ and $3^{\prime}$ quartiles) |  | P. laevis s.l.s intensity (median - 1' and 3' quartiles) |  |
|  |  |  | in intestinal tract | outside intestinal tract |  | Parasites in intestinal tract | Parasites outside intestinal tract | Parasites in intestinal tract | Parasites outside intestinal tract |
| Scardinius erythrophthalmus <br> (BPF) Rudd, Se | O | $6 / 4$ | 4 | 0 | 0.68 (0.26-0.94) | $1(0-2.25)$ | $0(0-0)$ | 1.5 (1-2.75) | - |
| Leuciscus leuciscus (BPF) Common Dace, Ll | V | $7 / 6$ | 6 | 0 | 0.86 (0.47-0.99) | $6(1-11)$ | $0(0-0)$ | $6.5(1-13)$ | - |
| Phoxinus phoxinus <br> (BPF) Minnow, Pp | O | $68 / 40$ | 18 | 31 | 0.24 (0.17-0.38) | $0(0-1)$ | $0(0-3)$ | $2(1-5.25)$ | $4(2-7)$ |
| Cypriniformes Balitonidae |  |  |  |  |  |  |  |  |  |
| Barbatula barbatula (BF) | O | 86 / 33 | 32 | 2 | 0.37 (0.28-0.48) | $1(0-1)$ | 0 (0-0) | 1,5 (1-2) | 1 |
| Loach, Bba <br> Siluriformes - <br> Ictaluridae |  |  |  |  |  |  |  |  |  |
| Ameiurus melas (BF) Catfish, Am | V | 6 / 6 | 2 | 5 | 0.18 (0.06-0.74) | 0 (0-3.25) | 2 (0.75-25.3) | 4.5 (2-7) | $2(1.5-27.5)$ |
| Perciformes - Percidae |  |  |  |  |  |  |  |  |  |
| Perca fluviatilis <br> (BPF) Perch, Pf | O | 16 / 10 | 10 | 0 | 0.63 (0.38-0.83 | $2(0-4.75)$ | $0(0-0)$ | 3.5 (2-6) | - |
| Scorpaeniformes Cottidae |  |  |  |  |  |  |  |  |  |
| Cottus gobio (BF) Bullhead, Cg | O | $30 / 10$ | 10 | 0 | 0.33 (0.18-0.51) | $0(0-1)$ | $0(0-0)$ | $1(1-1.5)$ | - |
| Gasterosteiformes Gasterosteidae |  |  |  |  |  |  |  |  |  |

Table 2: Parameters of a power function (Taylor's power law) fitting the relationship between the logarithms of variance in abundance and mean abundance, across fish species hosts of Pomphorhynchus laevis s.l. from two populations. Abundance was recorded for adult intestinal parasites. The parameters $a$ and $b$ are from the function $\operatorname{Var}(\mathrm{A})=\mathrm{a} \cdot \mathrm{M}(\mathrm{A})^{\mathrm{b}}$, with the variance in parasite abundance and mean abundance estimated for nine fish species for which at least 25 individuals were collected. The parameter $b$ represents an index of spatial heterogeneity (or aggregation) of parasite distribution across fish species. Goodness of fit is given by the coefficient of determination $r^{2}$, and the $P$-value. The slope (b) is significantly different from unity ( $\mathrm{F}_{1,6}=24.9, P=0.002$ )Bootsrapped values for the slope $b$ and $r^{2}$ and their 95\% intervals were calculated based on 1000 bootstrap replicates

| River | Intercept <br> (loga) | $b$ | Adjusted $r^{2}$ | $F$-test <br> statistics <br> $(\mathrm{df})$ | $P$ | \% var <br> explained |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Ouche | 0.74 | 1.66 | 0.96 | 157.4 | $<0.0001$ | 95.7 |
| $(\mathrm{~N}=9)$ |  | $[1.39-1.92]$ | $[0.80-0.98]$ | $(1,7)$ |  |  |

Table 3: Body condition metrics of fish infected with intestinal P. laevis s.l., (a) Body condition index (BCI) according parasite $\operatorname{load}(\log 10$ abundance), fish sex and their interaction, in the Ouche (OU) and Vingeanne (VI) rivers (see Fig. S6); (b) Hepathosomatic index (HIS) and gonadosomatic index (GSI), according to parasite load, fish species and sex.
(a)

$\left.\begin{array}{lcccc}\hline \text { Sex } & & \text { Chi2 }=4.34(4), & \text { Chi2 }=2.50(4), & \text { Chi2 }=5.55(4), \\ P=0.24\end{array}\right)$

## (b)

| HSI $(\mathrm{n}=246)$ | GSI ( $\mathrm{n}=256)$ |
| :--- | :--- |


| Log10 abundance | Barbel: Chi2 $=8.9$ (1), $P=0.003$ <br> Chub: Chi2 $=5.1$ (1),$P=0.024$ all others: non signif. | Chi2 $=7.92$ (9), $P=0.54$ |
| :---: | :---: | :---: |
| Fish species | / | Chi2 $=29.7$ (12), $P=0.003$ |
| Sex | Chi2 $=18.04$ (4), $P=0.0012$ | Chi2 $=77.83$ (4), $P<0.0001$ |
| Log10 abundance <br> * fish sp | Chi2 $=14.53$ (6), $P=0.02$ | Chi2 $=5.54$ (6), $P=0.48$ |
| $\log 10$ abundance * sex | Chi2 $=1.43$ (2), $P=0.49$ | Chi2 $=2.24$ (2), $P=0.20$ |

Supplementary material - Journal of Helminthology
Distribution of Pomphorhynchus laevis s.I. (Acanthocephala) among fish species at a local scale: importance of fish biomassdensity

Marie-Jeanne Perrot-Minnot, Loïc Bollache, Clément Lagrue

Supplementary Figure S1.


Fig. S1 Abundance distribution of intestinal P. laevis s.l. in different fish species sampled in the Ouche (OU) and Vingeanne rivers (VI), showing aggregate distributions of parasites among individual fish.

Supplementary Figure S2


Fig. S2. Relationship between Pomphorhynchus laevis s.l. abundance $(\log 10+1)$ and individual body size in nine fish species from two localities (OU: Ouche river; VI: Vingeanne river). For illustrative purpose, we categorized fish size in deciles (equal number of
individuals per category), except for gudgeon in the Vingeanne river (quintiles), using the package Hmisc v4.2.0 (Harrell, 2019) The results of fitting the abundance of $P$. laevis s.l. to fish size using log-linear relationship are reported as $P$-value and adjusted $\mathrm{R}^{2}$. Fish species for which sample size was low (<7) are not reported.

Harrell F. E. Jr, with contributions from Charles Dupont and many others. (2019). Hmisc:
Harrell Miscellaneous. R package version 4.2-0. https://CRAN.R-project.org/package=Hmisc

## Supplementary figure S3



Fig. S3. Mean intensity of intestinal Pomphorhynchus laevis s.l. in fish host from the (a) Ouche and (b) Vingeanne rivers. Mean intensity in fish species with different letters are significantly different, after post-hoc paired comparison (Dunn tests with B-Y method), those without letter are not significantly different from any other species.

Supplementary Figure S4


Fig. S4. Relationship between fish density (nb.m-2) and mean abundance of intestinal Pomphorhynchus laevis s.l. after log-transformation, across nine fish species from the Ouche River.

## Supplementary figure S5



Fig. S5. Relationship between parasite aggregation level (estimated using the variance-tomean ratio of abundance) and fish biomass-density, among nine fish species from the Ouche River.

Supplementary figure S6


Fig. S6. Relationship between body condition of individual fish and Pomphorhynchus laevis s.l. abundance, according to fish host sex and species. Body condition is measured as the residuals of the regression of whole body weight on body size ( $\log 10$-transformed).

Significant relationships are represented by a regression line. Numbers in bracket are sample sizes.

Supplementary figure S7


Fig. S6. Individual hepatosomatic index (HIS) in relation to (a) parasite load and (b) individual fish size, according to sex (female: plain line), for barbel and chub; and gonadosomatic index according to fish species (c) and sex (d) in the Ouche locality.

